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International Centre for Genetic Engineering and Biotechnology United Nations Industrial Development Organization



# **Collaborative Research Programme**

# TERMINAL EVALUATION REPORT

UNIDO contract # 91/049

ICGEB ref. #: CRP/CHI-90-01

**Project initiation:** 1991

**Project termination: 1995** 



International Centre for Genetic Engineering and Biotechnology United Nations Industrial Development Organization



## Collaborative Research Programme

## TERMINAL EVALUATION REPORT

Part 1

## **Title of Project**

Studies of the stress response in biomining microorganisms. Possible implications in the improvement of the bioleaching process.

Keywords: bioleaching, stress response, heat shock, starvation, Thiobacillus ferrooxidans.

UNIDO contract # 91/049	ICGEB ref. #: CRP/CHI-90-01
Project initiation: 1991	Project termination: 1995
Principal Investigator's name: Carlos A Affiliate Centre mail address :	A. Jerez
Departamento de Bioquímica, Facultad	de Medicina

Universidad de Chile, Casilla 70086, Santiago-7, Chile

physiological state in industrial operations.

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Fax no. (56 2) 73 563 84	Email address
Abstract:	

The biomining or bioleaching of minerals is a process by which metals such as copper, gold and uranium are solubilized from the ores containing them by the action of acidophilic microorganisms. During this process, bacteria are subjected to different kinds of environmental stresses such as temperature or pH changes and lack of essential nutrients that may affect the activity of the bacteria, therefore, limiting the efficiency and rate of metal extraction. In response to these stressing conditions, bacteria are known to change the genetic expression of several cellular polypeptides, inducing stress proteins and others as a defense or remediating mechanism to overcome the adverse condition.

The main objective of this project was to know the stress responses of extreme acidophiles such as *Thiobacillus ferrooxidans* at the molecular level. This is not only interesting from the fundamental point of view, but will help also to monitor the in situ state of these microorganisms, making possible to improve their activity in industrial processes.

Experimentally, we studied the global gene expression changes by using two-dimensional polyacrylamide gel electrophoresis of the proteins synthesized in vivo and identified some of the polypeptides induced by microsequencing of their N-terminal ends. We analyzed in detail the proteins synthesized by *T. ferrooxidans* when subjected to heat shock (from  $30^{\circ}$  to  $40^{\circ}$ C), cold shock (from  $30^{\circ}$  to  $10^{\circ}$ C), pH stress (from 1.5 to 3.5 and from 3.5 to 1.5) and the lack of nutrients such as phosphate or changes in nutrients, such as from ferrous iron to elementary sulfur, which usually take place in bioleaching operations. Some proteins were identified as stress proteins or molecular chaperones, and others as part of the different response systems. Based on the results obtained, we begun to develop immunological methods that will allow to monitor the planktonic and ore-attached bioleaching microorganisms and their in situ

## OBJECTIVES/METHODOLOGY (proposed at the time of the submission of the research proposal)

## **OBJECTIVES**

- 1. To know the components specifically induced in response to pH, temperature and nutrient changes.
- 2. To prepare either antibodies or nucleic acid probes de detect the components and the levels in which they are induced.
- 3. To assess the relative physiological condition of the bacteria in a given bioleaching operation (for example, if it is preferentially oxidizing sulfur or if starved for a given nutrient) by using the probes prepared. Decisions could then be made, if possible, to change the conditions to improve the local bacterial activity (e.g., add phosphate, etc).

## Methodology:

<u>Detection of proteins synthesized under stress conditions.</u> To study the global regulatory responses of *T. ferrooxidans*, we employed analysis of the proteins synthesized under different stressing conditions by means of two-dimensional polyacrylamide gel electrophoresis of both non-labeled and radio-labeled bacteria. The chemolithotrophic microorganisms were labeled by growing them in the presence of either Na<sub>2</sub><sup>14</sup>CO<sub>3</sub> or [<sup>35</sup>S]methionine.

<u>Analysis of some specific stress-induced proteins</u>. The localization of some of the proteins that changed their levels under different conditions was done by preparing outer membrane proteins or by <sup>125</sup>I-labeling in the presence of Iodogen. Identification of some of the proteins was done by employing immunological reactions. Some of the proteins of interest, were identified by microsequencing of their amino-terminal ends. The polypeptides were extracted from Commassie Blue stained 2-D gels (one or more spots of the same protein to have around 100 pmoles of each species). After rehydration, they were concentrated by running them in an SDS-PAGE. After this, the protein was transfered to a membrane of polyvinylidene difluoride (PVDF) for direct automatic Edman microsequencing.

<u>Preparation of polyclonal antibodies.</u> Polyclonal antibodies against whole bacterial cells of *T. ferrooxidans* were employed to monitor the microorganisms under different conditions. Polyclonal antibodies were also prepared against surface proteins induced by the lack of phosphate.

<u>Reverse genetics and expression of some proteins induced under different growth conditions.</u> To identify some of the genes of interest, whose expression varies during the different growth conditions, we started to employ reverse genetics. For this, we used the information generated from the N-terminal end sequences of the different proteins identified in 2-D gels. From these sequences, and the information of codon usage by *T. ferrooxidans* we synthesized the appropriate degenerate oligonucleotides. With this oligonucletides we generated the corresponding probes to identify and isolate the DNA fragments containing the genes in the genomic DNA from *T. ferrooxidans*.

RESULTS

## Compare against the set objectives)

<u>Heat shock response.</u> We have previously studied the heat shock response of mesophilic bioleaching microorganisms such as *Thiobacillus ferrooxidans* and *Leptospirillum ferrooxidans* and also of the thermophilic archaeon *Sulfolobus acidocaldarius*. We found that all these microorganisms apparently induced heat shock proteins. In *T. ferrooxidans* we found that the main proteins induced upon heat shock were the equivalents to *E. coli* DnaK and GroEL proteins (5, 13). This was established by immunological analysis (western blott) and by microsequencing of the N-terminal end of the corresponding proteins (5). These polypeptides showed a great identity with the *E. coli* chaperones (70% for GroEL and 80% with DnaK) and also had an oligomeric structure. GroEL proteins from both bacteria formed a 14-mer, whereas *E. coli* DnaK protein existed partially as a dimer and the *T. ferrooxidans* DnaK-equivalent showed only a monomeric nature under our experimental conditions (5). We also found that the levels of phosphorylation of DnaK and GroEL and other proteins greatly varied when *T. ferrooxidans* were subjected to heat shock and phosphate starvation. These results support the possible role of phosphorylation of chaperones in the sensing and regulation of the stress response in bacteria (Seeger, M., Osorio, G. and Jerez, C. A., submitted).

<u>pH stress response.</u> We found that a pH shift (3.5 to 1.5) also elicited a heat shock-like response in *T. ferrooxidans*. When *T. ferrooxidans* cells grown at pH 1.5 were shifted to pH 3.5, there were several changes in the general protein synthesis pattern, including a large stimulation of the synthesis of a 36,000 molecular weight protein (p36). The apparent low isoelectric point of p36, its location in the membrane fraction, and its cross-reaction with anti-OmpC from *Salmonella typhi* suggested that it may be a porin whose expression is regulated by extracellular pH (1).

<u>Phosphate-starvation response.</u> To cope with phosphate limitation, bacteria such as *E. coli* have evolved complex regulatory systems to assimilate phosphorous very efficiently. Phosphate regulation is complex, perhaps because phosphate is an essential nutrient. In the case of *T. ferrooxidans*, nothing was known about the proteins that may be involved in the phosphate scavenging and uptake mechanisms. We analyzed the molecular response of *T. ferrooxidans* to phosphate starvation. Cultivation of the bacteria in the absence of added phosphate not only reduced their growth rate and capacity to oxidize ferrous iron but induced a remarkable filamentation of the cells (7). Two-dimensional get electrophoresis revealed at least 25 proteins whose levels of synthesis were increased upon phosphate limitation, as well as some polypeptides that were exclusively synthesized under the starving condition (6-8). One of the proteins induced by the lack of phosphate was an acid phosphatase with a pH optimum of about 3.8, and a M.W. of 26 kDa, which was apparently located in the periplasm. The N-terminal end sequence of the protein showed some homology with the known sequence of *Lysobacter enzymogenes* alkaline phosphatase and the *Escherichia coli* alkaline and acid (pH 2.5) phosphates (7). The proteins induced by *T. ferrooxidans* upon phosphate starvation are probably part of a phosphate scavenging system utilized by the microorganisms in their natural environment, in which soluble forms of phosphorous could be limiting.

We found that phosphate starvation of *T. ferrooxidans* increased its capacity to attach to a sulfide mineral and to elemental sulfur (9). To establish if this effect was due to changes in the bacterial surface components, we analyzed the outer membrane proteins and the lipopolysaccharide (LPS) by employing SDS-PAGE and Western blotting with antisera against either whole cells or LPS from *T. ferrooxidans*. The amount of LPS in the cells starved for phosphate was about 25% higher when compared with control cells. However, only minor differences in the average size of the repeating units of the polymer were seen (9). These results, and the fact that several outer membrane proteins were induced in the phosphate-starved cells (4) suggest that not only LPS, but surface proteins and possibly hydrophobicity may play an important role in *T. ferrooxidans* attachment to minerals (9, 12).

Some of these polypeptides could be traced to the surface of the bacteria (4). One of these proteins was the *T. ferrooxidans* outer membrane protein (Omp40), whose level of expression was apparently regulated by the external pH and the concentration of phosphorus. The amino terminal sequence of Omp40 showed little identity with the *Escherichia coli* OmpC, OmpF or PhoE porins, but it was 38.5% identical to the outer membrane channel-forming protein NosA from *Pseudomonas stutzeri*, whose expression is also regulated environmentally (4).

We have prepared antibodies against whole cells of *T. ferrooxidans*, *L. ferrooxidans* and *T. thiooxidans* to monitor these cells under different conditions (2, 3, 11). We also prepared polyclonal antibodies against the outer membrane proteins induced under phosphate starvation that will be used for immunofluorescence to monitor the in situ state of bacterial cells in simulated industrial conditions (García, A., Varela, P. and Jerez, C. A. "Evolution"

of the planktonic and ore-attached populations of bioleaching microorganisms and determination of their in situ physiological state". Abstract book, 7th International Symposium on Microbial Ecology, Aug. 27-Sept 1°, Santos, Brazil, 1995)

#### Work plan and time schedule (originally envisaged)

#### **RESULTS** (continued...)

<u>Changes in global expression of *T. ferrooxidans* when grown in sulfur. There is great interest in studying the structure, expression and regulation of the genes involved in the oxidation of ferrous iron and sulfur, since they make *T. ferrooxidans* and other bioleaching bacteria industrially important. Several proteins involved in ferrous ion oxidation by *T. ferrooxidans* are known, including rusticyanin, cytochromes a and c and other proteins. On the other hand, the proteins involved in sulfur oxidation have not been studied in detail and their cellular localization and exact roles are still unknown.</u>

To identify some of the gene products involved in these oxidations, we have initiated the study of the changes in global gene expression in *T. ferrooxidans*, when the bacteria is grown on sulfur or iron. A few polypeptides were exclusively synthesized when the cells were grown in ferrous iron, and others were apparently synthesized only when *T. ferrooxidans* was grown in sulfur (10, 13).

By microsequencing of their N-terminal ends, and by specific Heme group staining, we have identified some of these proteins as cytochromes. In addition, we are preparing antibodies against some of the bacterial surface proteins that change their expression according to the environmental conditions. These antibodies will also allow us to monitor the in situ physiological state of the bioleaching microorganisms. This is important to establish, since due to the chemical and biological leaching of sulfides, elemental sulfur is generated which tends to coat mineral particles, interfering with the process.

#### Work plan and time schedule (originally envisaged)

- 1. <u>Studies on the synthesis of different stress proteins.</u> During the first and second years the plan contemplated the study of the response of *T. ferrooxidans* to stressing conditions of temperature, abrupt pH changes (in their growth range, between 1.5 and 3.5) and lack of the essential nutrient phosphate.
- 2. <u>Characterization of some stress proteins.</u> The plan contemplated during the three years the characterization of outer membrane proteins, periplasmic phosphatases and the molecular chaperones DnaK and GroEL.
- 3. <u>Studies simulating industrial conditions.</u> Studies with ores were planned during the three years to monitor the microorganisms in conditions similar to the actual bioleaching operations.

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- 4. <u>Isolation of some genes of interest</u>. These studies were planned for the second and third years, once information on the proteins and their identification was obtained.
- 5. <u>Preparation of antibodies and DNA probes</u>. Antibodies against different components from *T. ferrooxidans* were planned for the second and third years.
- 6. <u>Use of antibodies and probes to monitor bioleaching operations.</u> These studies were planned at the end of the proyect.

t landmarks, duration of individual tasks (use bar charts); evaluation criteria (publications, ts, services, training)					
		First year	Second year	Third year	
Dat	es of payments reception	July 91	Feb 93	Jul 94	
Acti	ivities:				
1.	Studies on the synthesis of different stress proteins.	****	****		
2.	<u>Characterization of some stress</u> proteins.	*****	****	****	
3.	Preparation of antibodies against total bacteria and surface components.	****		****	
4.	Microsequencing of N-terminal ends of different polypeptides	****	XXXXXXXXX	****	
5.	Generation of DNA probes for reverse genetics			****	
6.	Studies of monitoring bacteria grown in ores.			****	
7.	Publications (numbers as in the list enclosed).	1-5	6-9	10-15	

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In general, most of the initial objectives planned were developed as expected. The characterization of some of the genes coding for proteins induced by the lack of phosphate in T. ferrooxidans, such as PhoB and PhoR and those apparently involved in preferential oxidation of sulfur was initiated at the end of the project and it will be further developed in future research projects.

## NETWORKING

We initiated collaboration with the group of Dr. Héctor Torres from Instituto de Investigaciones en Ingeniería Genética y Biología Molecular (INGEBI), Buenos Aires, Argentina.

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We have had collaborative interaction with Dr. Tomás Vargas, from Facultad de Ciencias Físicas y Matemáticas, Universidad de Chile, specially for the studies involving ores and monitoring of microorganisms.

We collaborated with Dr. Ricardo Amils, from Universidad Autónoma de Madrid, since they visited our laboratory to learn the 2-D PAGE procedures we employ for the bioleaching microorganisms, and we learned pulse field gel electrophoresis technique in Spain.

We also had exchange with Dr. Börje Lindström, from the University of Umeä in Sweden where we performed the first microsequencing of the proteins studied.

#### PUBLICATIONS

- 1. Amaro, A.M., Chamorro, D., Seeger, M., Arredondo, R., Peirano, I. and Jerez, C.A. (1991) Effect of external pH perturbations on in vivo protein synthesis by the acidophilic bacterium *Thiobacillus ferrooxidans*. J. Bacteriol. 173: 910-915.
- 2. Jerez, C.A. and Arredondo, R. (1991) A sensitive immunological method to enumerate Leptospirillum ferrooxidans in the presence of Thiobacillus ferrooxidans. FEMS Microbiol. Lett. 78: 99-102.
- 3. Jerez, C.A. and Arredondo, R. (1991) Dot-immunobinding assay for specific and fast detection and enumeration of different bioleaching microorganisms. In Bioleaching: from molecular biology to industrial applications (Badilla, R. Vargas, T. and Herrera, L., eds.) Ed. Universitaria, pp. 89-94.
- 4. Jerer, C.A., Seeger, M. and Amaro, A.M. (1992) Phosphate starvation affects the synthesis of outer membrane proteins in *Thiobacillus ferrooxidans*. FEMS Microbiol. Lett. 98: 29-34.
- 5. Varela, P. and Jerez, C.A. (1992) Identification and characterization of GroEL and DnaK homologues in *Thiobacillus ferrooxidans*. FEMS Microbiol. Lett. 98: 149-154.
- 6. Seeger, M. and Jerez, C.A. (1992) The lack of phosphate affects global gene expression in *Thiobacillus* ferrooxidans. Geomicrobiol. J. 10: 227-237.
- 7. Seeger, M. and Jerez, C.A. (1993) Phosphate-starvation induced changes in *Thiobacillus ferrooxidans*. FEMS Microhiol. Lett. 108: 35-41.
- 8. Seeger, M. and Jerez, C.A. (1993) Response of *Thiobacillus ferrooxidans* to phosphate limitation. FEMS Microbiol. Revs. 11: 37-42.
- Amaro, A.M., Seeger, M., Arredondo, R., Moreno, M. and Jerez, C.A. (1993). The growth conditions affect *Thiobacillus ferrooxidans* attachment to solids. In Biohydrometallurgical Technologies, Vol II. (A.E. Torma, M.L. Apel and C.L. Brierley, eds.). The Minerals, Metals & Materials Society pp. 577-585.
- Osorio, G., Varela, P., Arredondo, R., Seeger, M. Amaro, A.M. and Jerez, C.A. (1993) Changes in global gene expression of *Thiobacillus ferrooxidans* when grown in elemental sulfu. In Biohydrometallurgical Technologies, Vol II. (A.E. Torma, M.L. Apel and C.L. Brierley, eds.). The Minerals, Metals & Materials Society pp. 565-575.
- 11. Amaro, A. M., Hallberg, K. B., Lindström, E. B. and Jerez, C. A. (1994) An immunological assay for the detection and enumeration of thermophilic biomining microorganisms. Appl. Environ. Microbiol. 60: 3470-3473.
- 12. Arredondo, R., García, A. and Jerez, C. A. (1994) The partial removal of lipopolysaccharide from *Thiobacillus ferrooxidans* affects its attachment to solids. Appl. Environ. Microbiol. 60: 2846-2851.
- 13. Jerez, C. A., Varela, P., Osorio, G., Seeger, M., Amaro, A. M. and Toledo, H. Differential gene expression of *Thiobacillus ferrooxidans* under different environmental conditions. Minerals Bioprocessing II. (R. W. Smith, D. Holmes, eds.). The Minerals, Metals & Materials Society. In press.

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#### STATEMENT OF EXPENDITURES

To be filled by ICGEB Budgets as per original proposal		To be filled by the Affiliated Centre Summary of expenditures *		
2) consumables	US <b>\$</b>	2) consumables	US\$ 8,263.58	
3) training	US <b>\$</b>	3) training	US\$	
4) literature	US\$	4) literature	US\$ .1,433.22	
5) miscellaneous	US <b>\$</b>	5) miscellaneous	US\$	
TOTAL GRANT	US\$	TOTAL	us\$.10,000.00	

#### Please itemize the following budget categories (if applicable)

## Capital equipment

-Microcentrifuge US\$ 3,727.43

-Olympus fluorescence microscope and photomicrography attachments US\$ 16,635.42

-Power supply and electrophoresis chamber US\$ 2,561.65

-Laser printer US\$ 2,161.88

-Gel dryer, UV transilluminator, magnetic stirrer and photographic system US\$ 9,504.05

#### Training (provide names, duration of training, host laboratory)

a) Carlos A. Jerez attended as Organizer and Convener of the Coloquium "Molecular Biology and Biochemistry of Acidophilic Chemolithotrophs: Applications on Bacterial Leaching of Ores" and the additional presentation of a poster, American Society for Microbiology General Meeting, New Orleans, USA, 26-30 May, 1992.
b) Presentation of a poster by C. A. Jerez in the symposium "Advances in Modern Biotechnology, La Habana, Cuba, 8-12 June, 1992. Both events US\$ 3,000

Presentation of part of the work done in the project by Carlos A. Jerez in:

International Biohydrometallurgy Symposium, Jackson Hole, Wyoming, U.S.A., August 22-25, 1993. US\$ 2,646.60. 7th International Congress of Bacteriology and Applied Microbiology, Prague, Czech Republic, July 3-8, 1994. Partial financing of US\$ 600

#### Literature

-Geomicrobiology Journal, Special issue: Geomicrobial Agents and Processes for Bioleaching Metal Sulfide Ores. -Ausubel et al., Short Protocols in Molecular Biology, 1992, John Wiley & Sons, Inc.

-Charackliss and Marshall, Biofilms, 1990, John Wiley & Sons, Inc.

-Bergey's Manual of Determinative Bacteriology, 1994, Williams and Wilkins.

-Watson et al. Recombinant DNA, 1992. W. H. Freeman and Company and other specialized texts.

-Subscription to Molecular Microbiology, Applied and Environmental Microbiology and other specialized journals,

\* Please do not send involces, receipts etc.; these should be kept by the Affiliated Centre for future reference and sent to ICGEB upon request.

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UNIDO	STATEMENT OF ACCOUNTS	DATE : AUGUST 25 1995
CONTRACT	: Nº 31 / C49	
INSTITUTION	: UNIVERSIDAD DE CHILE	
PROYECT TITLE	: STUDIES OF THE STRESS RESPONSE IN FIO POSSIBLE IMPLICATIONS IN THE IMPROVE PROCESS.	
PROYECTS LEADE	R : DR. CARLOS A. FEREZ G.	-
REPORT FOR THE	PERIOD STARTING : FEERUARY 25 1994	ENDING : FEERUARY 25 1998 YEAR 3
INCOME: U.S.	\$. 10.000,00	
UNIDO FUNDS RE INTEREST EARNE	CEIVED (DATE) : JULY 28 1994 D : 0	

TOTAL INCOME. U.E.S. 19.00100

## EXPENDITURE

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POOR QUALITY



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We certify that the above expenditures were employed to support the research planned in UNIDO Contract N<sup>o</sup> 91/049, in agreement with UNIDO guidelines and the allowed items

Dr. CARLOS JEREZ Proyect leader.

F§.

Srta. 80 BARAHONA Financial Officer



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UNIDO	STATEMENT OF ACCOUNTS	DATE : JUNE 16 1994
CONTRACT	: Nº 91 / 049	
INSTITUTION	: UNIVERSIDAD DE CHILE	
PROYECT TITLE	: STUDIES OF THE STRESS RESPONSE IN BIOMINI POSSIBLE IMPLICATIONS IN THE IMPROVEMENT	
PROYECTS LEADER	: DR. CARLOS A. JERE?	
REPORT FOR THE PERIO	DD STARTING : FEBRUARY 25 1993	ENDING : FEBRUARY 25 1994.
INCOME: U.S.\$.	35.000,00	
UNIDO FUNDS RECEIVEL INTEREST EARNED : C	) ( DATE ) : FEBRUARY 16 1993 )	

TOTAL INCOME: U.S.S. 35,000,00

EXPENDITURE

ITEN	ALLOCATED IN US.\$.	RECEIVED IN US.\$.	EXPENDITURE IN US.\$.	ITEM BALANCE BROUGHT FND.
SMALL EQUIPEMENT Microscopio Gastos de Aduana	14.000,00	14.000,00	15.635,42 12.512,00 4.123,42	(2.635,42)
<i>CONSUMABLES &amp; CHEMICALS Chemicals Miscellaneous</i>	13.000,00	13.000,00	13.401,55 6.587,23 6.814,32	(401,55)
EDUCATION & TRAINING (travel and per diem) Airplane Tickets Stay Expenses	4.000,00	4.000,00	3.246,60 1.521,00 1.725,60	753,40
LITERATURE Books	1.000,00	1.000,00	669,65 669,65	330,35
OTHER EXPENSES Inscrip. Syzposium	3.000,00	3.000,00	1.046,43 1.046,43	1.953,57
TOTAL	35.000,00	35.000,00	34.999,65	0,35

# UNIVERSIDAD DE CHILE - FACULTAD DE MEDICINA - DIVISION DE CIENCIAS MEDICAS NORTE

## DEPARTAMENTO DE BIOQUINICA

Independencia N° 1027 Block C. 1-2 Teléfonos: 370081-776560 Anexos: 5270-5271-5272 Dirección Postal: Casilla 70086 Santiago - 7 CHILE

UNITED NATIONS INDUSTRIAL DEVELOPMENT ORGANIZATION

UNIDO CONTRACT Nº 91/049

REFERENCE Nº CRP/CH190-01)

#### INANCIAL STATEMENT

FOR THE PERIOD 07/91 TO 07/92

NAME OF PRINCIPAL INVESTIGATOR: CARLOS JEREZ GUEVARA.

INSTITUTION: UNIVERSIDAD DE CHILE

PROJECT TITLE: Studies of the stress response in biomining microorganisms. Possible implications in the improvement of the bioleaching process.

AMOUNT RECEIVED FOR PERIOD 07/91 TO 07/92 US\$ 35.000.-

BUDGET CATEGORY

11

 SMALL EQUIPMENT
 18.000. 

 CONSUMABLES & CHEMICALS
 10.000. 

 EDUCATION & TRAINING
 3.000. 

 LITERATURE
 1.000. 

OTHER EXPENSES \_\_\_\_\_\_

TOTAL EXPENDITURES 35.000.-

FINANCIAL OFFICER: FERNANDO NAVAS M. (Signature) (print) PHONE: 7370081/5213 DATE: 10/05/92