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ESTABLISHMENT OF AN INTERNATIONAL

LACTIC ACID FERMENTATION TECHNOLOGY NETWORK

(LABNET)

bу

K. KOMAGATA - Consulton +

Redisdep Eff. 45 Comphell

ESTABLISHMENT OF AN INTERNATIONAL LACTIC ACID FERMENTATION TECHNOLOGY NETWORK [LABNet]

We are all aware of the fact that biotechnology and genetic engineering can have a unique potential to improve disadvantaged populations. However, a technology assessment is often very difficult since the socio-economic environment can have a profound impact on its success. When the focus is on profits and efficiency, results of technology assessment will be quite different from the one that will emerge if the focus is on sustainable development and employment generation. The reason for this problem lies mainly with the economist, who at present is ab'e to evaluate an industry defined by

products but has great difficulties with an industry which is a *means* of production.

In recent years, the achievements of modern biotechnology and genetic engineering have been associated with a rapid growth of networks in bioinformatics, and they are now becoming extremely popular in the developed world. So far, however, the emphasis was restricted mainly to new drugs and improved seeds for the major food crops. The synergistic potential between communication technology and modern biotechnology in the agroindustrial area has only recently started and was pioneered by the Biofocus Foundation, which underlined the potential of genetic engineering for the upgrading of traditional technologies by means of an improvement management of microbial starter cultures.

Lactic acid bacteria have been chosen initially because of their importance for the manufacture of fermented foods and dairy products as well as their impact on health care all over the world. Initially it is vital to concentrate on the importance of starter cultures on fermented dairy products.

Milk is the normal habitat of a number of lactic acid bacteria which may cause spontaneous souring. For a sophisticated control, however, modern industrial processes utilize specially prepared lactic acid bacteria as starter cultures in the manufacture of fermented dairy products. A starter usually consists of

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harmless microorganisms which, upon culturing in milk or milk-based mixes, impart desirable and predictable characteristics of flavour and texture attributable to a certain fermented milk product. A multi-strain starter is considered to have an advantage over a single strain starter since fermentation will continue in the presence of a phage which specifically attacks one strain only. In Australia and New Zealand, the preferred technology is to grow two single starters individually and to mix them prior to culturing of cheese vats. The advantage of this technique is that an individual strain, preselected for its performance and resistance to antibiotics and phages, is used for ensuring greater predictability and uniformity of culture effects. A wealth of information on the starter cultures in dairy products is available now in the literature.

For the distribution of starters an earlier method involved shipping them as a liquid culture containing about 10° organisms/ml. However, subculturing caused loss of critical characteristics. Presently, the most common method for distribution is in the form of frozen concentrates containing approximately 10¹¹ organisms/ml. The culture concentrates are standardized for activity by the culture manufacturer and has eliminated the need for routine maintenance at a plant level. The starters are graded according to their acid production, rate of growth, phage and antibiotic resistance, and ability to develop

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typical flavour and texture.

The production of lactic acid, acetic acid, CO₂, diacetyl and acetaldehyde from lactose and citric acid is of fundamental importance to the growth of lactic cultures and generation of characteristic flavour in products. The fermentation reactions are carried out by homo- and heterofermentative systems. Furthermore, improvements in the desired product formation can easily be achieved through genetic modification.

The starter is therefore the most crucial component, and it is essential to develop and maintain appropriate microbiological expertise in the propagation, maintenance, and control of lactic starter cultures. Culture propagation should neverthless still be conducted in a specified, secluded area of the plant where access of personnel is restricted. Airborne contamination and phage infection must be controlled.

Control of acid and flavour development in lactic acid starters cultures may be achieved by understanding their growth characteristics. By modifying the inoculation rate, incubation temperature, and time, it is possible to direct the fermentation, in a limited way, to fit the plant schedules.

In short, the microbiological activities of starter cultures should control the growth of spoilage bacteria and pathogens. Latest developments in microbial process development has shown that the success of most microbial

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technologies hinges on the state and condition of the starter culture or inoculum.

Lactic acid bacteria starter cultures are, however, not only used in dairy products, but are more important,

i.e. in Southeast Asia in fermented foods, fermented beverages, vegetable fermentations, fodder preparations (silage), and the utilization of agricultural wastes, manure, slaughterhouse waste and fishery waste.

Furthermore, lactic acid bacteria play a very significant role in human intestinal flora and thus exhibit a number of health effects.

Against this background of the vast potential of lactic acid bacteria to influence the life-quality in developing countries, a natural setting for a research and development effort ought to be given high priority.

Mutation and the various mechanisms for DNA transfer have undoubtedly resulted in a reservoir of naturally occurring strains of lactic streptococci which vary considerably in genotype. There has also been recently much speculation about genetical engineering of starter cultures. The first concern must be to provide a starter culture that is resistant to phage and still has a high rate of acid production. Little is known, however, about the basic biology of phage resistance in lactic acid bacteria.

It is only in the last couple of years that the full potential of lactic acid bacteria has started to be fully

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realized in the form of isolated genes, plasmid vectors, chromosomal vectors, chromosomal integration vectors and I have just heard a few genetical approaches at the 9th International Biotechnology Symposium in Washington in August this year. These presentations, however, were mainly concerned with modifications of the carbohydrate metabolism to enhance flavour production.

For the practical applications of genetic engineering, metabolic stability, 'food-grade' genetic systems and phage resistance are all important goals.

The overall goal therefore of the global network is to strengthen the local scientific and technological capabilities in developing countries and to do this in a network fashion, where members in poor countries are supported by their colleagues in the industrialized world. The network would be designed to facilitate international cooperation where the members from developing countries can identify their needs and solve their own problems.

It is proposed to develop a non-profit but self-supporting international network on fermentations by lactic acid bacteria over a five year period. The aims is a decentralized structure designed to achieve an action-oriented synergistic interaction between specialised activities, largely based on established laboratory networks. LABNet would have four major functions:

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- the screening for methods, strains and applications that will be of special significance to developing countries;
- 2. open information exchange on research activities related to the impact of lactic acid bacteria on health and nutrition in man and animals, on food technology and on the upgrading of agricultural residues and of fish- and slaughterhouse wastes;
- management of expert task forces set up to achieve specific results;
- 4. support of entrepreneural activities and indigenous creativity in the lactic acid bacteria field.

It is visualized that the international network will be organized from a combination of small face-to-face meetings and computer-assisted conferences. Action-oriented' working groups and innovative 'task forces' will communicate via computer conferences with the help of expert moderators. The overall project will be coordinated by a project manager with a small staff supplemented with fellowship workers being trained in communication technology, data base use and starter culture management.

In order to avoid 'duplication' and 'reinventing the wheel', LABNet will seek in the long-term close cooperation with existing international organizations and partnerships with scientific organizations, especially MIRCENS. •.•

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For the launching of LABNet it is important that its aims, approaches and progress are made widely known through partners that can act as information channels, for example existing networks such as the various information networks.

Bangkok MIRCEN

REVIEW: CURRENT STATE

Scope of the Bangkok MIRCEN

The Microbiological Resources Center for the Southeast Asia (Bangkok MIRCEN) was established in 1976 at the Thailand Institute of Scientific and Technological Research (TISTR), Bangkok, Thailand. The first major task of Bangkok MIRCEN was to identify areas of activities of regional economic development where microorganisms play key roles. A survey was made on culture resources in collections in Thailand and in other ASEAN member countries, namely, Indonesia, Malaysia, Philippines, and Singapore. It was found from this survey that the R & D activities were more or less confined to food fermentation and other industrial fermentation. Technical information related to cultures, particularly their potential utilization, was lacking, and information exchanges in this field were limited to personal communications.

Bangkok MIRCEN serves the microbiological community of the region with the following functions:

1) Collection, preservation, characterization, and distribution of microorganisms --bacteria and fungi including yeasts-- important for biotechnology, education, and applied

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research.

2) Collection and dissemination of information relevant to the cultures and their application.

3) Promotion of research activities directed towards the needs of the origin.

4) Training of manpower and fellowship program.

5) Acting as a liaison.

Financial, Personnel, and Facilities Supports

Financial supports for Bangkok MIRCEN has largely been contributed by the Government of Thailand, through TISTR, in the form of technical personnel, research funds, and other supporting facilities. Contribution of the Government of Thailand is approximately accounted for US\$ 95,000 a year. One director, three full-time persons, two part-time persons, and a technician, and the personnel expenses are provided by TISTR. They are dealing with management and service business on the collection and the maintenance of the microbial cultures. Additional funds have been provided by UNESCO, UNEP, ASM, and other sources. Major necessary equipment, chemicals, electricity, gas, and water supply are provided by TISTR.

Culture Collection Activities

Bangkok MIRCEN maintains a collection of microorganisms important for agriculture and industry. It has approximately 2,500 strains of bacteria, yeasts, molds, and algae in 1991. A majority of these microorganisms are preserved in freezedried form.

Since Bangkok MIRCEN started the operation, it has distributed approximately 4,500 cultures to universities and researc institutions within Thailand and approximately 400 cultures to other countries. Attempts have also been made to acquire economically important cultures from collections in the region for deposition at Bangkok MIRCEN. During its 16year operation, approximately 1,700 cultures have been obtained from several institutions of different countries.

Catalogues of holdings of Bangkok MIRCEN have been prepared in 1975, 1979, 1985, 1987, and 1990, and a (atalogue of the 4th edition contains over 1,300 strains of bacteria, yeasts, molds, and algae. Bangkok MIRCEN has prepared lists of cultures and other publications relevant to culture collections and biotechnology, and these publications have been widely distributed.

Bangkok MIRCEN has been responsible for the assembly of information on microbiological institutions in the Southeast Asian region and data on collections of microorganisms, and Thailand Directory of Collections of Cultures of Microorganisms was published in 1988, and the microbial data are stored by the use of dBASE III plus.

Co-operating Laboratories

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Bangkok MIRCEN has, at present, 4 co-operating laboratories in Hong Kong, Indonesia, Malaysia, and Philippines. These laboratories interact with Bangkok MIRCEN in the promotion of MIRCEN activities at the national level and cooperate in dissemination of information. Bangkok MIRCEN helps strengthen their roles by providing technical assistance especially in the field of culture collections, funding for maintenance of culture collections and research grants. Bangkok MIRCEN maintains close links with its cooperating laboratories.

Workshop and Training Courses

Regional workshops and trainings in the field of applied microbiology including culture collections have been one of the main activities of Bangkok MIRCEN. These workshops and training courses have been organized periodically at national and as well as regional levels. The ultimate objective of the training activities is to strengthen regional and local capabilities in the development and maintenance of culture collections. In addition, an intensive on-the-job training on culture collections has been provided on request. A total of 666 trainees have participated in the training program. Of this number, 21 trainees were from China, Hong Kong, Indonesia, Laos, Malaysia, Pakistan, Philippines, Trinidad, and UK.

Research Grant

Research grants within the framework of the MIRCEN program have been available for Bangkok MIRCEN to disburse on behalf of the MIRCEN to collaborating laboratories and researchers of the region.

Research Activity

Bangkok MIRCEN has been actively engaged in research activities in the areas of the MIRCEN framework. Funds have been made available for foreign scientists and professors to carry out joint cooperative research programs at the Bangkok MIRCEN and vice versa. A program aiming at upgrading efficiency and widening the scope of usefulness of microorganisms for industrial and agricultural applications is also carried out. The work includes the isolation and testing of new strains as well as strains in Bangkok MIRCEN Culture Collection.

Regional and International Collaboration

Bangkok MIRCEN has collaborated with several regional and international bodies by participating in and contributing to regional and international workshops, training courses, meetings, conferences, and other activities.

Promotional Activities

A number of short articles on Bangkok MIRCEN and its ac-

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tivities (in Thai and English) have been regularly published in several science journals for the purpose of familiarizing scientists and students to become aware of the beneficial roles of microorganisms and the information of the source on the subject. Bangkok MIRCEN has also issued newsletters and other useful publications for distribution to other MIRCENs, educational and research institutions within Thailand and abroad.

ASSESSMENT

Bangkok MIRCEN has contributed through activities of its culture collection and research in enhancing IISTR's visibility and competence in the fields of applied and environmental microbiology and biotechnology. Bangkok MIRCEN Culture Collection is a medium-sized collection, and manpower and equipment are not fully satisfied. However, Bangkok MIRCEN Culture Collection has maintained approximately 2,500 microbial strains, and these strains are rather well-documented by the use of personal computers. Approximately 4,500 strains have been distributed to universities, institutions, and commercial firms not only in Thailand but also foreign countries. Speciality of Bangkok MIRCEN Culture Collection is its holdings of microbial strains, namely, cultures in this collection are consisted

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of isolates from sources in Thailand, and the collection is not a copy of other culture collections. Particularly, algal collection is unique because a small number collections are maintaining algal strains in the world.

Bangkok MIRCEN has been serving as a key organization in the fields of applied and environmental microbiology and biotechnology in the Southeast Asian region, and conducting a lot of cooperating research projects with Thai researchers and with those in neighboring countries. This cooperation is useful for technology transfer and exchange of information. The development of technology is now being transferred to rural community in Thailand. In connection with the cooperation, Bangkok MIRCEN has been conducting a lot of workshops, training courses, and on-the-jot trainings relevant to culture collections and biotechnology. Trainees jointed in these meetings are accounted for 666, and they have come from Thailand and foreign countries. This organization for such meetings should be highly evaluated for development of applied and environmental microbiology and biotechnology in the Southeast Asian region.

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LIST OF MIRCEN DIRECTORS

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Professor A. KORNHAUSER, Biotechnological Information Exchange System (BITES) MIRCEN, Unesco International Centre for Chemical Studies, P.O.Box 18/1, 61001 Ljubljana, Yugoslavia.

Professor S.T. CHANG, Bioconversion Technology MIRCEN, Department of Applied Biology, The Chinese University of Hong Kong, Shatin, N.T., Hong Kong.

Participants: Université de Technologie (Compiègne), INSBANA (Dijon), Centre d'Immunologie INSERM-CNRS (Marseille), Laboratoire de Chimie bactérienne CNRS (Marseille), INRA (Montpellier), Institut Pasteur (Paris), INSA (Toulouse).

THAILAND'S LABNET NODE

UNIDO'S INTERNATIONAL LABNET NETWORK

A Proposed Resource List

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by

K. Komagata

THAILAND'S LACTIC ACID BACTERIA COLLECTION

AT BANGKOK MIRCEN

Microbiological Service Unit National Center of Genetic Engineering for Biotechnology

at

Thailand Institute of Scientific and Technological Research Bangkok Thailand 1994 Microbiological Resources Center (Bangkok MIRCEN)

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Thailand Institute o. Scientific and Technological Research (TISTR)

STAFF MEMBERS

Director		Poonsook Atthasampunna
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	Bacteria	Wanchern Potacharoen
	Fungi	Suparp Artjariyasripong
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		Vullapa Arunpairojana
		Suntud Sirianantapaiboon
		Bundit Fungsin

Culture collection of lactic acid bacteria in Thailand at the Bangkok MIRCEN began few years ago under the research project on isolation and screening of microorganisms play important role in fermented food products. There are two types of cultures in the coolection. One is the reference strains and another is the local isolates. Cultures are preserved by freeze-dired method in which regular viability check are carried out in the quality control of cultures. Number of total strains held in the collection at prestent is 156 strains of which is 50 working reference strains and 106 local isolates. Most of reference strains have been obtained from Japan and ATCC.

The cultures are distributed on request with free charge on the exchange basis and minimal charge for the governmental officers or students. The regular charge is for the private company and foreign country. The culture fee are:

250 Bath for governmental officers and students500 Bath for private company25 US\$ for the foreign country

The collection accepts deposition of cultures of known genus and species with emphasis to the specific characters which are important in education, research or industry. The deposition fee is free for the non-restricted cultures but charge for the restricted one at a variable rate set up at each condition.

The cultures are listed alphabetically in the first part of the catalogue and numerically in the second one. The sequence of the data supplied is:

- 1. Scientific names of the species arranged in alphabetical order, followed by authorities for the scientific names.
- 2. TISTR accession number, followed by equivalent designation in other collections and/or by the name and strain designation of the isolator.
- 3. History of the strain or name of the depositor.
- 4. Source of the isolate.
- 5. Special applications of the strain, when known, are listed together with a reference number in brackets.

ABBREVIATIONS

ATCC	American Type Culture Collection, Rockville, Maryland,
	U.S.A.
DMKU	Department of Microbiology, Faculty of Science, Kasetsart
	University, Bangkok, Thailand.
DSM	Deutsche Sammlung von Mikroorganismen, Gottingen, Germany.
LAM	Institute of Applied Microbiology, University of Tokyo, Japan.
JCM	Japan Collection of Microorganisms, RIKEN, Wako, Saitama,
	Japan.
MSDS	Microbiology Section, Department of Science Energy, Bangkok,
	Thailand.
NISL	Noda Industrial Science Laboratory, Noda, Japan.
MIST	National Institute of Science and Technology, Manila,
	Philippines.
NRIC	NODAI Research Institute Culture Collection, Tokyo University
	of Agriculture, Tokyo, Japan.
NRRL	Northern Utilization Research and Development Division, U.S.
	Department of Agriculture, Peoria, Illinois, U.S.A.
TUA	Department of Agricultural Chemistry, Tokyo University of
	Agriculture, Tokyo, Japan
UPPC	University of the Phillippines Culture Collection, Diliman,
	Quezon City, Phillipines.
UQM	Microbiology Department, University of Queensland, Brisbane,
	Australia.

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Lactobacillus acidophilus (Moro) Hansen & Mocquot

388	M.Kozaki <-TUA 002			
	Homofermentative Lactic acid produced	95.83	%	[01]

450 M.Kozaki <-TUA 346L

Lactobacillus brevis

855	S. Tanasupawat A314 [O2] =NRIC 0130 SOURCE : Fermented fish spawn (<i>Kai-pla dong</i>)
860	S. Tanasupawat P46-1 [02] =NRIC 0134 SOURCE : Pickled bastard mustard (<i>Pak-sian dong</i>)
868	S. Tanasupawat P28-3 [02] =NRIC 0137 SOURCE : Fermented bamboo shoot (<i>Naw-mai dong</i>)
Lactobacillus	buchneri (Henneberg) Bergey et al.

048 DMKU <-B.Yongsmith <-ATCC 4005 =ATCC 4005 SOURCE : Tomato pulp

Lactobacillus bulgaricus (Orla-Jensen) Rogosa & Hansen

451 M.Kozaki <-TUA 093L

Lactobacillus casei (Orla-Jensen) Rogosa & Hansen

389	M.Kozaki <-TUA 016			
	Homofermentative			
	Lactic acid produced	91.06	%	[01]

390 Bagasses SOURCE : Bagasses Homofermentative Lactic acid produced 90.90 %

453 M.Kozaki <-TUA 164L

Lactobacillus casei subsp. rhamnosus Rogosa et al.

- 047 DMKU <-B.Yongsmith <-ATCC 7469 [03] =ATCC 7469 Type strain Assay of riboflavin
- 372 M.Kozaki <-TUA 333

Lactobacillus cellobiosus Rogosa et al.

W. Daengsubha 7 [01] 398 SOURCE : Bagasses Heterofermentative Lactic acid produced 40.45 % Grow at 45 C

Lactobacillus confusus

881	S. Tanasupawat (P322-1) [03] =NRIC 0130 SOURCE : Rice wine (Satoh-dong)
934	S. Tanasupawat <-NRIC 1544 =NRIC 1544 =ATCC 27646 [02 ;03] SOURCE : Lettuce leaves

Lactobacillus curvatus

938	S. Tanasupawat
	=NRIC 1052 =ATCC 25601
	SOURCE : Milk
	%G+C 42.9

Lactobacillus delbrueckii (Leichmann) Beijerinck

108	pl. pathology res. dept. <-NRRL B-445 Homofermentative Lactic acid produced 87.35 % [01]
326	UPCC 77 <-NIST

326 =NRRL B-763

Lactobacillus delbrueckii subsp. lactis (Orla-Jensen) Weiss et al.

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785	DMKU <-ATCC 7830 [03]
	=ATCC 7830
	Assay of cobalamin

Lactobacillus fermentum Beijerinck

DMKU <-ATCC 14931 =ATCC 14931	[03]
SOURCE : Fermented	beets
	DMKU <-ATCC 14931 =ATCC 14931 SOURCE : Fermented Neotype

- W. Daengsubha 1 [01] 391 SOURCE : Bagasses Heterofermentative Lactic acid produced 41.79 % Grow at 45 C
- ATCC 9338 [03] 683 Assay of amino acids

879	S. Tanasupawat (P1-35) [02] =NRIC 0132 SOURCE : Pickled black mustard (<i>Fak-gard dong</i>)
914	S. Tanasupawat F10-2 [02] =NRIC 0135 SOURCE : Fermented bean sprout (<i>Tua-ngok dong</i>)
915	S. Tanasupawat FN12-1 [02] =NRIC 0129 SOURCE : Fermented fish (<i>Pla-chao</i>)
937	S. Tanasupawat F26-1 SOURCE : Fermented fish (<i>Pla-som</i>) %G+C 51.4
945	S. Tanasupawat KM7-1 SOURCE : Fermented rice noodle (<i>Khanom-jeen</i>)
946	S. Tanasupawat KM17-1 SOURCE : Sour pork sausage (<i>Sai-krok prieo</i>)
947	S. Tanasupawat KM26-1 SOURCE : Fermented rice flour (<i>Khanom-jeen</i>)
948	S. Tanasupawat KM27 SOURCE : Fermented rice flour (<i>Khanom-jeen</i>)
949	S. Tanasupawat KM12 SOURCE : Pickled bastard mustard (<i>Pak-sian dong</i>)
950	S. Tanasupawat KM24-1 SOURCE : Fermented rice flour (<i>Khanom-jeen</i>)

Lactobacillus halotolerans

939	Sausage [03]
	=DSM 20190 ;ATCC 35410
	Type strain
	%G+C 44.8

Lactobacillus lactis (Orla-Jensen) Bergey et al.

452 M.Kozaki <-TUA 026L

Lactobacillus leichmannii (Henneberg) Bergey et al.

- 449 UQM 1364
- 476 MSDS <-ATCC 7830 [Lactobacillus delbrueckii] Assay of vitamin B12 %G+C 50.5

Lactobacillus mali

919	S. Tanasupawat	<-NRIC	1076T		
	=ATCC 27053 SOURCE : Apple Type strain	juice,	Cider	press	[03]

Lactobacillus pentosus

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845	S. Tanasupawat (A35) =NRIC 1827 SOURCE : Sour pork sausage (<i>Sai-krok prieo</i>) % G+C 46
847	S. Tanasupawat FN2 SOURCE : Fermented fish (<i>Pla-som</i>) %G+C 47
848	S. Tanasupawat FN4 SOURCE : Fermented pork (<i>Nham</i>) %G+C 45
849	S. Tanasupawat FN35 SOURCE : Fermented pork (<i>Nham</i>)
851	S. Tanasupawat F36-1 SOURCE : Fermented fish (<i>Pla-som</i>) %G+C 45
852	S. Tanasupawat F37-2 SOURCE : Fermented fish (<i>Pla-som</i>)
853	S. Tanasupawat F31-5 =JCM 8333 SOURCE : Fish cake (<i>Som-fak</i>) %G+C 46
856	S. Tanasupawat A83 SOURCE : Fermented fish (<i>Pla-som</i>)
857	S. Tanasurawat P7-1 =NRIC 1831 ;JCM 8336 SOURCE : Pickled black mustard (<i>Pak-gard dong</i>) %G+C 45
859	S. Tanasupawat P24-1 SOURCE : Pickled bastard mustard (<i>Pak-sian dong</i>) %G+C 46
865	S. Tanasupawat F5-1S SOURCE : Pickled onion (Hom-dong) %G+C 47
867	S. Tanasupawat F21-1 =JCM 8337 SOURCE : Fermented bamboo shoot (<i>Naw-mai dong</i>) %G+C 45
869	S. Tanasupawat F32-1 SOURCE : Fermented bamboo shoot (<i>Naw-mai dong</i>) %G+C 47
870	S. Tanasupawat A330 =NRIC 1836 SOURCE : Spoiled durian %G+C 46

- FP2-1 871 S. Tanasupawat SOURCE : Fermented tea leaves (Miang) %G+C 15 S. Tanasupawat FP18 872 SOURCE : Fermented tea leaves (Miang) %G+C 47 S. Tanasupawat FP38-1 873 SOURCE : Fermented tea leaves (Miang) S. Tanasupawat FP46-2 874 SOURCE : Fermented tea leaves (Miang) %G+C 47 S. Tanasupawat SM01 878 =NRIC 1839 SOURCE : Fish cake (Som-fak) S. Tanasupawat F11-1 880 SOURCE : Pickled bastard mustard (Pak-sian dong) %G+C 46 920 S. Tanasupawat =NRIC 1069T <-ATCC 8041 SOURCE : Corn Silage Type strain %G+C 46 S. Tanasupawat Al 940 =NRIC 1551 SOURCE : Fermented tea leaves (Miang) A4 S. Tanasupawat 941 =NRIC 1552
- SOURCE : Fermented tea leaves (Miang)

Lactobacillus plantarum (Orla-Jensen) Bergey et al.

DMKU <-ATCC 8014 050 Assay of amino acids %G+C 45.1 373 M.Kozaki <-TUA 354 Requirement for pantothenic acid DMKU <-P.Srisomwong Ll 541 SOURCE : Fermented pork (Nham) Homofermentative Grow at 15 C ,45 C DMKU <-P. Srisomwong L9 542 SOURCE : Fermented pork (Nham) Homofermentative Grow at 15 C ,45C

543	DMKU <-P.Srisomwong L33 SOURCE : Fermented pork (<i>Nham</i>) Strong acid production Grow at 45 C
544	DMKU <-P. Srisomwong L35 SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 15 C ,45 C
844	S. Tanasupawat F7-1 =JCM 8341 SOURCE : Sour pork sausage (<i>Sai-krok prieo</i>) %G +C 44
846	S. Tanasupawat F33-1 SOURCE : Sour beef sausage [Mum] %G+C 43
850	S. Tanasupawat FP48-1 =NRIC 1829 SOURCE : Sour beef sausage [Mum] %G+C 44
854	S. Tanasupawat F35-1 SOURCE : Fermented fish (<i>Som-fak</i>) %G+C 45
858	S. Tanasupawat P16 SOURCE : (<i>Pak-nam dong</i>) %G+C 44
861	S. Tanasupawat P30-1 =NRIC 1832 =JCM 8344 SOURCE : (Pak-koom dong) %G+C 44
862	S. Tanasupawat F10-1 =JCM 8346 SOURCE : Pickled bean sprouts (<i>Tau-ngok dong</i>) %G+C 44
863	S. Tanasupawat P17 SOURCE : Sauerkraut %G+C 43
864	S. Tanasupawat F2-1 SOURCE : Pickled onion (Hom-dong) %G+C 45
926	S. Tanasupawat KM1-1 SOURCE : Sweeted rice (<i>Khao-mak</i>) %G+C 44
951	S. Tanasupawat SOURCE : Fermented rice flour (Khanom-jeen)

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Lactobacillus sake

890	S. Tanasupawat FN1-4 =NRIC 0125 SOURCE : Fermented pork (<i>Nham</i>)
911	S. Tanasupawat F3-1 =NRIC 0126 SOURCE : Fermented pork (<i>Nham</i>)
912	S. Tanasupawat =NRIC 0128 SOURCE : Fermented fish (<i>Som-fak</i>)

Lactobacillus sp.

539	DMKU <-P. Srisomwong L37 SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 15 C
540	DMKU <-P. Srisomwong L39 SOURCE : Fermented pork (<i>Nham</i>) Heterofermentative Grow at 15 C ,45 C
866	S. Tanasupawat F20 =NRIC 0136 SOURCE : Pickled onion (<i>Hom-dong</i>)
882	S. Tanasupawat FP37-1 SOURCE : Fermented tea leaves (<i>Miang</i>) % G+C 47
891	S. Tanasupawat A28 =NRIC 0127 SOURCE : Fermented pork (<i>Nham</i>)
913	S. Tanasupawat FP51-1 =NRIC 0133 SOURCE : (<i>Pak-koom dong</i>)

Leuconostoc dextranicum (Beijerinck) Hucker & Pederson

056 DMKU <-ATCC 19255	056	DMKU	<-ATCC	19255
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- 377 M.Kozaki <-TUA 204
- 454 M.Kozaki <-TUA 204L
- 474 DMKU <-B.Saeng-on 8m8/25B SOURCE : Sugar factory

Leuconostoc mesenteroides (Tsenkovskii) van Tieghem

053 DMKU <-ATCC 10830 Production of dextran

- 120 UPCC 44
- 473 DMKU <-B. Saeng-on 8m8/25A SOURCE : Sugar factory Production of dextran
- 478 MSDS <-ATCC 8042 Assay of amino acids
- 942 S. Tanasupawat =NRIC 1541 ;ATCC 8293 Type strain

Pediococcus acidilactici Lindner

- 051 DMKU <-ATCC 8042 Assay of amino acids
- 397 NISL 7113
- 424 DMKU <-S. Tanasupawat N53 [06;07;08] SOURCE : Fermented pork (Nham)
- 425 DMKU <-S. Tanasupawat N54 [06;07;08] SOURCE : Fermented pork (Nham)
- 783 ATCC 33314 [03] Type strain

Pediococcus halophilus Mees

332	DMKU B-119 <-S. Suknaisilp no.78 SOURCE : Soy sauce
333	DMKU B-120 <-S. Suknaisilp no.82 SOURCE : Soy sauce Lactic acid produced 88.24 % [09]
334	DMKU B-121 <-S. Suknaisilp no.83 SOURCE : Soy sauce Lactic acid produced 97.96 % [09]
429	DMKU <-S. Tanasupawat PH01 SOURCE : Fermented fish (<i>Pla-ra</i>) Lactic acid produced 97.96 % [09]
430	DMKU <-S. Tanasupawat PH25 [10] SOURCE : Fermented fish (<i>Pla-ra</i>)
431	DMKU <-S. Tanasupawat PH27 [10] SOURCE : Fermented fish (<i>Pla-jom</i>)
432	DMKU <-S. Tanasupawat PH45 [10] SOURCE : Fermented fish extract (<i>Nam-pla</i>) Lactic acid produced 93.65 %

- 433 DMKU <-S. Tanasupawat PH47 [10] SOURCE : Fermented sea clam (Hoi-dong)
- 434 DMKU <-S. Tanasupawat PH59 [10] SOURCE : Fermented shrimp (Kung-jom)
- 435 DMKU <-S. Tanasupawat PH65 [10] SOURCE : Fermented fish (Pla-jom)
- 436 DMKU <-S. Tanasupawat PH88 [10] =JCM 2016 SOURCE : Fermented saltwater fish (*Bu-du*)
- 437 DMKU <-S. Tanasupawat PH120 [10] SOURCE : Fermented fish entrails (Tai-pla)
- 438 DMKU <-S. Tanasupawat PH130 [10] SOURCE : Fermented fish entrails (Tai-pla)
- 439 DMKU <-S. Tanasupawat PH155 [10] SOURCE : Fermented fish (*Pla-jom*)

Pediococcus pentosaceus Mees

374 TUA P-19

- 413 DMKU <-S. Tanasupawat NO1 [07;10] SOURCE : Fermented pork (Nham) Lactic acid produced 91.14 %
- 414 DMKU <-S. Tanasupawat N31 [07;10] =JCM 2023 SOURCE : Fermented fish cake (Pla-som) Lactic acid produced 84.72 % %G+C 31
- 415 DMKU <-S. Tanasupawat N37 [07;10] =JCM 2024 SOURCE : Pickled bastard mustard (*Pak-sian dong*) Lactic acid produced 96.98 % %G+C 37.5
- 416 DMKU <-S. Tanasupawat N38 [07;10] SOURCE : Fermented pork (Nham) Grow at 42 C
- 417 DMKU <-S. Tanasupawat N78 [07;10] SOURCE : Fermented pork (Nham) Lactic acid produced 82.01 %
- 418 DMKU <-S.Tanasupawat N91 [10] SOURCE : Pickled black mustard (*Pak-gard dong*)
- 419 DMKU <-S. Tanasupawat N133 [07;10] =JCM 2027 SOURCE : Fermented pork sausage (Sai-krok prieo)

420	DMKU <-S. Tanasupawat N256 [07 ;10] SOURCE : Fermented pork (<i>Nham</i>) Lactic acid produced 90.17 %
421	DMKU <-S. Tanasupawat N271 [07 ;10] SOURCE : Fermented fish cake (<i>Pla-som</i>) Lactic acid produced 96.65 %
422	DMKU <-S. Tanasupawat N278 [07 ;10] SOURCE : Fermented pork sausage (<i>Sai-krok prieo</i>) Grow at 42 C
423	DMKU <-S. Tanasupawat N295 [07 ;10] SOURCE : Fermented fish cake (<i>Som-fak</i>) Grow at 42 C
954	DMKU <-S. Tanasupawat SOURCE : Fermented rice (<i>Khao-mak</i>)
955	DMKU <-S. Tanasupawat SOURCE : Fermented rice (<i>Khao-mak</i>)

Pediococcus sp.

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129	M.Sundhagul SOURCE : Japanese fermented food
394	DMKU <-S. Tanasupawat <-IAM1684
529	DMKU <-P. Srisomwong P1 [05] SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 45 C
530	DMKU <-P. Srisomwong P8 [05] SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 45 C
531	DMKU <-P. Srisomwong P2 [05] SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 45 C
532	DMKU <-P. Srisomwong P3 [05] SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 45 C
533	DMKU <-P. Srisomwong P5 [05 ;06] SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 45 C
534	DMKU <-P. Srisomwong P10 [05 ;06] SOURCE : Fermented pork (<i>Nham</i>) Homofermentative Grow at 45 C

- 535 DMKU <-P. Srisomwong P11 [05] SOURCE : Fermented pork (*Nham*) Homofermentative Grow at 45 C
- 536 DMKU <-P. Srisomwong P17 [05;06] SOURCE : Fermented pork (Nham) Homofermentative Grow at 45 C
- 537 DMKU <-P. Srisomwong P18 [05;06] SOURCE : Fermented pork (Nham)
- 538 DMKU <-P. Srisomwong P20 [05;06] SOURCE : Fermented pork (*Nham*) Homofermentative Grow at 45 C
- 545 DMKU <-P. Srisomwong P27 [05] SOURCE : Fermented pork (Nham) Homofermentative Grow at 45 C
- 546 DMKU <-P. Srisomwong P31 [05] SOURCE : Fermented pork (*Nham*) Homofermentative Grow at 45 C

Pediococcus urinae-equi Mees

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426	DMKU <-S. Tanasupawat N86 [07] =JCM 2021 SOURCE : Pickled black mustard (<i>Pak-gard dong</i>) Lactic acid produced 84.90 %
427	DMKU <-S. Tanasupawat UO1 [07] SOURCE : Horse urine Lactic acid produced 88.33 %

Streptococcus cremoris Orla-Jensen

- 058 W. Daengsubha SOURCE : Yoghurt starter
- 456 M.Kozaki <-TUA 439L

Streptococcus faecalis Andrewes & Horder

- 379 M.Kozaki <-TUA 369 =ATCC 19433 [Enterococcus faecalis] =ATCC 19433
- 459 M.Kozaki <-TUA 194L

Streptococcus lactis (Lister) Lohnis

457 M.Kozaki <-TUA 154L

Streptococcus sp.

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Streptococcus	thermophilus Orla-Jensen
933	S. Tanasupawat SOURCE : Fermented beef sausage [Mum]
932	S. Tanasupawat SOURCE : Fermented tea leaves (<i>Miang</i>)
931	S. Tanasupawat SOURCE : Pickled black mustard (<i>Pak-gard dong</i>)
930	S. Tanasupawat SOURCE : Fermented pork sausage (<i>Sai-krok prie</i> o)
923	S. Tanasupawat SOURCE : Fermented fish (Pla-som)
928	S. Tanasupawat SOURCE : Fermented fish (<i>Pla-jom</i>)

458 M.Kozaki <-TUA 196L

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047	Lactobacillus casei subsp. rhamnosus
048	Lactobacillus buchneri
050	Lactobacillus plantarum
051	Pediococcus acidilactici
053	Leuconostoc mesenteroides
055	Lactobacillus fermentum
056	Leuconostoc dextranicum
058	Streptococcus cremoris
108	Lactobacillus delbrueckii
120	Leuconostoc mesenteroides
129	Pediococcus sp.
326	Lactobacillus delbrueckii
332	Pediococcus halophilus
333	Pediococcus halophilus
334	Pediococcus halophilus
372	Lactobacillus casei subsp. rhamnosus
373	Lactobacillus plantarum
374	Pediococcus pentosaceus
377	Leuconostoc dextranicum
379	Streptococcus faecalis
388	Lactobacillus acidophilus
389	Lactobacillus casei
390	Lactobacillus casei
391	Lactobacillus fermentum
394	Pediococcus sp.
397	Pediococcus acidilactici
398	Lactobacillus cellobiosus
413	Pediococcus pentosaceus
414	Pediococcus pentosaceus
415	Pediococcus pentosaceus
416	Pediococcus pentosaceus
417	Pediococcus pentosaceus
418	Pediococcus pentosaceus
419	Pediococcus pentosaceus
420	Pediococcus pentosaceus
421	Pediococcus pentosaceus
422	Pediococcus pentosaceus
423	Pediococcus pentosaceus

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424	Pediococcus acidilactici
425	Pediococcus acidilactici
426	Pediococcus urinae-equi
427	Pediococcus urinae-equi
129	Pediococcus halophilus
430	Pediococcus halophilus
431	Pediococcus halophilus
432	Pediococcus halophilus
433	Pediococcus halophilus
434	Pediococcus halophilus
435	Pediococcus halophilus
436	Pediococcus halophilus
437	Pediococcus halophilus
438	Pediococcus halophilus
439	Pediococcus halophilus
449	Lactobacillus leichmannii
450	Lactobacillus acidophilus
451	Lactobacillus bulgaricus
452	Lactobacillus lactis
453	Lactobacillus casei
454	Leuconostoc dextranicum
456	Streptococcus cremoris
457	Streptococcus lactis
458	Streptococcus thermophilus
459	Streptococcus faecalis
473	Leuconostoc mesenteroides
474	Leuconostoc dextranicum
476	Lactobacillus leichmannii
478	Leuconostoc mesenteroides
529	Pediococcus sp.
530	Pediococcus sp.
531	Pediococcus sp.
532	Pediococcus sp.
533	Pediococcus sp.
534	Pediococcus sp.
535	Pediococcus sp.
536	Pediococcus sp.
537	Pediococcus sp.

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538	Pediococcus sp.
539	Lactobacillus sp.
540	Lactobacillus sp.
541	Lactobacillus plantarum
542	Lactobacillus plantarum
543	Lactobacillus plantarum
544	Lactobacillus plantarum
545	Pediococcus sp.
546	Pediococcus sp.
683	Lactobacillus fermentum
783	Pediococcus acidilactici
785	Lactobacillus delbrueckii subsp. lactis
814	Lactobacillus plantarum
845	Lactobacillus pentosus
846	Lactobacillus plantarum
847	Lactobacillus pentosus
848	Lactobacillus pentosus
849	Lactobacillus pentosus
850	Lactobacillus plantarum
851	Lactobacillus pentosus
852	Lactobacillus pentosus
853	Lactobacillus pentosus
854	Lactobacillus plantarum
855	Lactobacillus brevis
856	Lactobacillus pentosus
857	Lactobacillus pentosus
858	Lactobacillus plantarum
859	Lactobacillus pentosus
860	Lactobacillus brevis
861	Lactobacillus plantarum
862	Lactobacillus plantarum
863	Lactobacillus plantarum
864	Lactobacillus plantarum
865	Lactobacillus pentosus
866	Lactobacillus sp.
867	Lactobacillus pentosus
868	Lactobacillus brevis
869	Lactobacillus pentosus

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870	Lactobacillus pentosus
871	Lactobacillus pentosus
872	Lactobacillus pentosus
873	Lactobacillus pentosus
874	Lactobacillus pentosus
878	Lactobacillus pentosus
87.9	Lactobacillus fermentum
880	Lactobacillus pentosus
881	Lactobacillus confusus
882	Lactobacillus sp.
890	Lactobacillus sake
891	Lactobacillus sp.
911	Lactobacillus sake
912	Lactobacillus sake
913	Lactobacillus sp.
914	Lactobacillus fermentum
915	Lactobacillus fermentum
919	Lactobacillus mali
920	Lactobacillus pentosus
926	Lactobacillus plantarum
928	Streptococcus sp.
929	Streptococcus sp.
930	Streptococcus sp.
931	Streptococcus sp.
932	Streptococcus sp.
933	Streptococcus sp.
934	Lactobacillus confusus
937	Lactobacillus fermentum
938	Lactobacillus curvatus
939	Lactobacillus halotolerans
940	Lactobacillus pentosus
941	Lactobacillus pentosus
942	Leuconostoc mesenteroides
945	Lactobacillus fermentum
946	Lactobacillus fermentum
947	Lactobacillus fermentum
948	Lactobacillus fermentum
949	Lactobacillus fermentum

950	Lactobacillus fermentum
951	Lactobacillus plantarum
954	Pediococcus pentosaceus
955	Pediococcus pentosaceus

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REFERENCES

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01	Daengsubha, W., T. Toyota, S. Okada, and M. Kozoki.
	1980. Taxonomic study of some lactic acid bacteria isolated
	from bagasses. Annual Reports of ICME, 3: 252-255.
02	Tanasupawat, S., S. Okada, K-I Suzuki, M. Kozaki and
	K. Komagata. 1993. Lactic acid bacteria, particularly
	heterofermentative lactobacilli found in fermented foods in
	Thailand. Bull. JFCC vol. 9 : 65-78.
03	American Type Culture Collection Staff (ed.). 1982.
	Catalogue of Strains I. 15th Ed. American Type Culture
	Collection, Rockville, Maryland, U.S.A. 755 pp.
04	Tanasupawat, S., T. Ezaki, K-I. Suzuki, S. Okada, K. Komagata
	and M. Kozaki. 1992. Characterization and Identification of
	Lactobacillus pentosus and Lactobacillus plantarum Strains
	from Fermented Foods in Thailand. J. Gen. Appl. Microbiol.,
	38, 121-134.
05	Srisomwong, P. 1985. The study of lactic acid bacteria in
	Nham. Progress report submitted to Assoc. Prof. K.A. Bukle,
	School of Food Technology, Faculty of Applied Science,
	University of South Wales, Sydney, Australia.
06	Utarapichat, 0. 1987. Selection of Salmonella-inhibiting
	lactic acid bacteria and preparation of powder inoculum of
	Nham. Master's Thesis (in Thai). Department of Microbiology,
	Regular of Science Kogeteent University Randkok Theiland.
	Faculty of Science, Raselsart University, Dangkok, Inatiand.
	124 pp.

07 Tanasupawat, S., and W. Daengsubha. 1983. *Pediococcus* species and related materials in Thailand. J. Gen. Appl. Microbiol. 29: 487-506.

- Tanasupawat, S., and K. Komagata. 1988. DNA base composition 80 of strains in Pediococcus species. Bullectin of the Japan Federation for Culture Collections 4: 63-67.
- Ueda, K. 1980. Search and screening of microorganisms having 09 decolorizing activity of molasses pigment. JSPS-NRCT Seminar on Agro-Industry Including Microbial Technology, Osaka, Japan; Microbial Utilization of Renewable Resources 1: 195-198.
- Tanasupawat, S. 1981. A taxonomic study of tetradgrouping 19 bacteria. Master's Thesis (in Thai), Department of Microbiology, Faculty of Science, Kasetsart University, Bangkok, Thailand. 285 pp.