



**TOGETHER**  
*for a sustainable future*

## OCCASION

This publication has been made available to the public on the occasion of the 50<sup>th</sup> anniversary of the United Nations Industrial Development Organisation.



**TOGETHER**  
*for a sustainable future*

## DISCLAIMER

This document has been produced without formal United Nations editing. The designations employed and the presentation of the material in this document do not imply the expression of any opinion whatsoever on the part of the Secretariat of the United Nations Industrial Development Organization (UNIDO) concerning the legal status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries, or its economic system or degree of development. Designations such as "developed", "industrialized" and "developing" are intended for statistical convenience and do not necessarily express a judgment about the stage reached by a particular country or area in the development process. Mention of firm names or commercial products does not constitute an endorsement by UNIDO.

## FAIR USE POLICY

Any part of this publication may be quoted and referenced for educational and research purposes without additional permission from UNIDO. However, those who make use of quoting and referencing this publication are requested to follow the Fair Use Policy of giving due credit to UNIDO.

## CONTACT

Please contact [publications@unido.org](mailto:publications@unido.org) for further information concerning UNIDO publications.

For more information about UNIDO, please visit us at [www.unido.org](http://www.unido.org)



# D00424

United Nations Industrial Development Organization

DISTRIBUTION  
LIMITED

ID/WG. 34/54  
23 July 1969

ORIGINAL: ENGLISH

International Petrochemical Symposium on the  
Development of the Petrochemical Industries in  
Developing Countries

1969, TOKYO, 27 - 31 October 1969  
**21**

PET SYM. / 14

## CHEMICALS FROM HYDROCARBONS BY THE FERMENTATION PROCESS

by

Shukue Kincshita and Takeo Suzuki  
Tokyo Research Laboratory  
Kyowa Hakko Kogyo Co. Ltd.  
Machida  
Tokyo

<sup>1)</sup> The views and opinions expressed in this paper are those of the author and do not necessarily reflect the views of the secretariat of UNIDO.  
This document has been reproduced without formal editing.



Distribution  
Library

PL/MR.34/54/PUB. 17  
23 July 1970

ORIGIN: ENGLISH

## United Nations Industrial Development Organization

Interregional Petrochemical Symposium on the  
Development of the Petrochemical Industries  
in Developing Countries

PEI. SYM. 3/12

Kiev, USSR, 20 - 31 October 1969

### SUMMARY

#### CHEMICALS FROM HYDROCARBONS BY THE FERMENTATION PROCESS 1/

by

S. Kinoshita  
T. Suzuki

Kyowa Hakko Kogyo Company Limited  
Tokyo, Japan

Microbial utilization of petroleum hydrocarbons have been studied in at least four directions in Japan. The first of these is the production of single cell protein, the second is the replacement of the raw materials for fermentation by hydrocarbon or the secondary products derived from petroleum, the third is the production of useful substances characteristic in hydrocarbon fermentation, and the fourth is the chemical modification of petroleum hydrocarbons by means of the oxidative activity of microorganisms.

Recently, some of techniques established initially on the laboratory scale have been developed on the semi-industrial level. For example, with yeast cell protein on a reasonable prospect, there is potential for full industrial operation in near future.

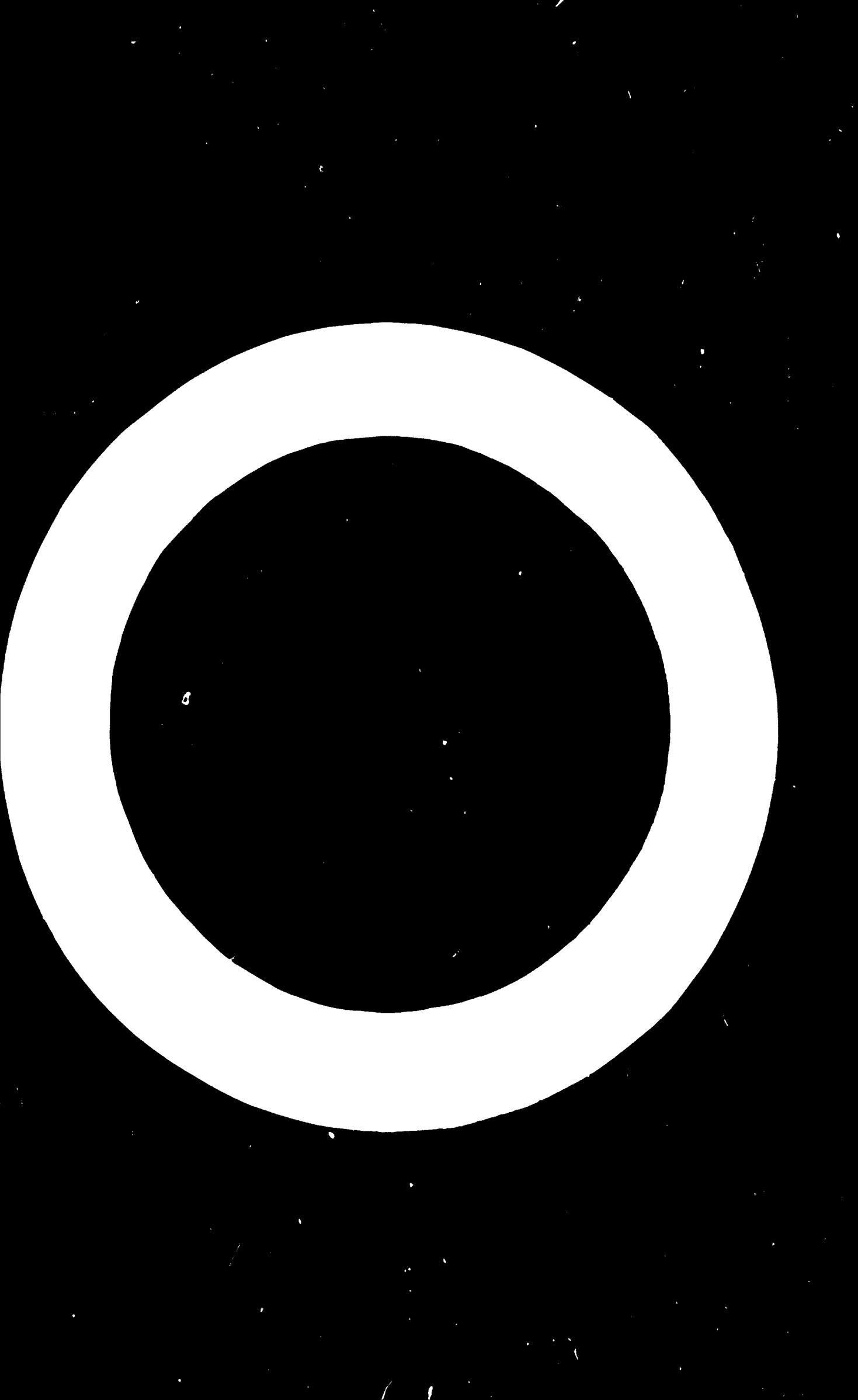
1/ The views and opinions expressed in this paper are those of the authors and do not necessarily reflect the views of the secretariat of UNIDC. This document has been reproduced without formal editing.

In connection with the fermentation processes of amino acids, particularly glutamic acid which was first established in Japan, the possibility is foreseen as the replacement of the raw materials by n-paraffin. Moreover, the production of threonine, norleucine, ornithine, citrulline and other amino acids from hydrocarbon have become possible owing to the success in obtaining a variety of bacterial mutants.

The production of sugars such as glucose, galactose, mannose and polysaccharides by bacteria and yeast grown on n-paraffin as the sole source of carbon have recently reported. Biochemical studies on the hydrocarbon fermentation have demonstrated the occurrence of considerable amount of glycolipids characteristic in hydrocarbon-utilizing bacteria.

Among the organic acids producible from hydrocarbon by microorganisms, the most promising product may be citric acid. The reported yield is more than 80 g/l from 60 g/l of n-paraffin.

In addition, the production of various chemicals such as vitamins, coenzymes, aromatic compounds and others has been reported in Japan. Many other oxidative products of aromatic hydrocarbon have been demonstrated.



### Introduction

With the improvement of the microbe-utilizing techniques, the fermentation process concerning the production of various useful chemicals has been developed to industrial scale, and further a great deal of the raw material has been demanded with it. However, it is now predicted that the supply of more sufficient amount of the raw material which has been so far obtained from agricultural products become more difficult in future because the shortage of the food in the world is foreseen with the rapid increase in the world population. And again, together with the trouble that agricultural product is not supplied constantly by year on account of various natural calamities, an increasing attention has been paid towards the replacement of the raw material of the fermentation by petroleum hydrocarbon which is estimated to be semi-exhaustively contained in the earth.

Considering the possible origin of petroleum on the basis of the scientific evidences, it is likely that organic matter such as proto-petroleum had already existed upon the earth before the living systems were generated. On the other hand, the present evolutionary theory suggests us that the most ancient living organisms might be the microbes capable of growing upon the organic matter which had already been formed on the earth. Therefore, the microbes and petroleum could probably have acted on each other since that period. The discovery that a number of microorganisms capable of utilizing hydrocarbon as the sole source of carbon are distributed widely in the soil is quite reasonable if the above consideration would be acceptable.

The tendency towards the replacement of the raw material by petroleum hydrocarbon appear to be an adverse current when considering from the evolutionary view point of living systems, but rather more extensive possibility could be expected in the attempt for the production of useful material.

In the process of conventional fermentation, it is known that the variation of the metabolic system and the product occurs whenever different substrate is supplied as carbon source. Therefore, more extensive variation in the replacement of the raw material by hydrocarbon is expectable.

Microbial conversion of hydrocarbon to useful products has been investi-

gated in at least four directions in Japan. The first of these is to produce protein more efficiently by hydrocarbon-utilizing yeast. At the international conference on single cell protein in 1967, any developed countries demonstrated that the techniques had been established on the semi-industrial scale. A second direction is to replace the raw material in the fermentation process by petroleum hydrocarbon and to produce the chemicals which have been already manufactured through the conventional processes. Actually, amino acids such as glutamic acid, threonine, homoserine, ornithine, citrulline and others, organic acids such as citric and  $\alpha$ -keto glutaric acid, and vitamins have been obtained by this process. A third direction for the study on hydrocarbon fermentation is on the basis of the assumption that various useful chemicals characteristic in the substrate hydrocarbon might be obtained. The production of fatty acids, alcohols, waxes and other lipids, sugars, and aromatic compounds have been reported. A fourth direction is concerned with the chemical modification of hydrocarbon, in which a part conversion of substrate hydrocarbon by microbial oxidation has been undertaken. Namely, the conversion of n-paraffin into the corresponding dicarboxylic acid by diterinal oxidation, alicyclic hydrocarbon into the corresponding alcohol and ketone, xylene into toluic acid, and cymene into eunic acid have been shown.

Besides the above-mentioned attempts, the materials derived secondarily from petroleum, which will be easily supplied with the recent growth of petrochemical industry, have been applied for fermentation processes as the raw materials. For example, glutamic acid is actually producible from ethanol or acetic acid with good yields.

The purpose of this communication is to review briefly Japanese studies concerning the production of the chemicals from hydrocarbon through the fermentation processes which have been attempted in the above-mentioned directions.

### 1. Hydrocarbon-utilizing microorganisms

Since there are many excellent publications<sup>1,2/</sup> concerning microbial reaction on hydrocarbon, it is not intended here to refer exhaustively to reported findings. The recent data obtained along with the investigation of the production of amino acids, sugars and other chemicals will be described in this paper.

It has been shown that the microbes capable of utilizing hydrocarbon include not only bacteria but also mold, actinomycetes and yeast. Those microbes are found widely in the soil regardless of the presence of hydrocarbon. Therefore, it is not so difficult to select the microbes capable of producing useful products from hydrocarbons. The growth of those microbes is initiated by the supply of aqueous nutrients and vitamins in addition to hydrocarbon as a carbon source under the optimal physical conditions. The most preferable carbon source for their growth is n-paraffins in the range of C<sub>11</sub> - C<sub>20</sub> (3-5). The optimum range of carbon number of n-alkane is different according to the by microbial strain used.<sup>5)</sup> For instance, some strains of Arthrobacter, Brevibacterium and Pseudomonas grow preferably on n-paraffin of C-15 to C-18, while some of Corynebacterium prefer those of C-12 to C-14. A similar tendency is also observed in the reductivity of amino acids and organic acids. Aromatic hydrocarbons<sup>6)</sup> or phenol compounds<sup>7)</sup> are generally less utilized by microorganisms.

On the other hand, a number of hydrocarbon-utilizing bacteria is also capable of growth on alcohols<sup>8)</sup>, organic acids or saccharides as the sole source of carbon. The ability of the growth on alcohols suggests some possibility of the utilization of alcohols and hydrocarbons. In this case, that of methanol or ethanol is a more preferable carbon source. Similarly, the fact that the carboxylic acids which are the intermediate in hydrocarbon oxidation (which is natural to them) as far as it is known, these bacteria are able to grow on carbohydrates as well, for example, on sugar alcohol such as mannitol, sorbitol and glycerol, and ketohexose such as fructose, galactose etc., although not preferable carbon sources<sup>8)</sup>. The characteristics of hydrocarbon-utilizing bacteria in sugar utilization could become a significant one in an attempt to produce sugars from hydrocarbon.

Another feature of hydrocarbon-utilizing microbes is that the demand of oxygen for their growth is higher than with others<sup>9)</sup>. In an attempt to produce a useful material from hydrocarbon through the fermentation process, the supply of sufficient oxygen should be considered in the growing phase of microbes. Some oxygen-containing products are derived from hydrocarbons which do not contain oxygen, hydrocarbon fermentation could be referred to as oxygen

<sup>5)</sup> Lactic acid is also utilized by some of them.

fixing fermentation. The additional effect of metal ions such as iron, copper and others could be explained on the basis of the above findings.

Of the physical conditions for the culture of bacteria, the most important factor is to control pH of culture medium below 0.5, because these bacteria are generally susceptible to death on higher pH and productivity is also depressed under such condition.

Further, the fact that these bacteria can take up both the nutrients of different nature, lipophilic carbon sources and hydrophilic other nutrients, suggests the presence of some special mechanisms characteristic of their cells. It is generally observed that those bacteria display a significant activity of emulsion formation in the mixture of hydrocarbons and aqueous solution of other nutrients. From these observations, we assume that a surfactant-like substance should be present on the surface of their cells and attempted to characterize it. Consequently, the isolated material was identified as trehalose ester of a hydroxy fatty acid.<sup>10/</sup> The glycolipids placed under the same category as the trehalose lipid have also been found in other microbial strains which can utilize n-alkane as the sole source of carbon, and the isolated materials displayed remarkable activity as the surfactant. These findings strengthen the possibility that these lipid play a possible role in the utilization of hydrocarbon by bacteria. In addition, occurrence of sophorose lipid in n-paraffin-utilizing yeast was also reported. Comparison of the characteristics of hydrocarbon-utilizing bacteria with those of others is of great interest to either bacteriologist or biochemist.

## 2. Production of amino acid from n-alkane

Owing to great progress in the fermentation techniques since the direct process of glutamic acid production was invented in Japan, L-amino acid has been supplied smoothly to various fields, the demand of amino acid has been rapidly increasing not only for food and feed supplement but also for the raw material of synthetic fibers and leathers. On these backgrounds, the replacement of the raw materials for fermentation by cheaper and more constantly supplied materials has been desired, and investigated for about seven years in Japan.

Early in the study, the question was to find the bacteria having both the characters which are capable of growing on n-alkane and also to convert it efficiently to glutamic acid or other amino acids. Fortunately, a number of microbial strains which are producible various amino acids have been discovered. Iizuka et al.<sup>12)</sup> first reported the occurrence of microbes in the soil, which were able to produce glutamic acids, alanine, tyrosine and lysine when kerosene was used as the sole source of carbon. And they<sup>13)</sup> named them as Corynebacterium hydrocarboelastus and C. oleophilus. In spite of that these microbes utilized n-alkane actively, the quantity of amino acids produced was not sufficient enough to develop to a further process. Subsequently, Yamada and Takahashi<sup>14)</sup> isolated the bacteria belonging to Corynebacterium, Arthrobacter, and Bacillus capable of producing various amino acids. A variety of microorganisms capable of converting kerosene and n-paraffin efficiently into glutamic acid and  $\alpha$ -ketoglutaric acid has been isolated from the soil by Tanaka et al.<sup>5)</sup> in our laboratory. These were classified to genus Corynebacterium, Arthrobacter, Hycopacterium, Brevibacterium, Micrococcus, Facillus and Noardia. Iguchi et al.<sup>15)</sup> have also shown that the bacterial strain isolated by them produced relatively large amounts of glutamic acid in the culture medium. Almost all of these bacteria have been known to require thiamine essential for the growth. It is of interest to compare this with the action of biotin which is known as the growth factor of the Corynebacterium glutamicus and is responsible for glutamic acid production from carbohydrate source.

#### Glutamic acid

Yamada and his colleagues<sup>16)</sup> have demonstrated using Corynebacterium hydrocarboelastus to yield 5 g/l of glutamic acid from n-paraffin under the suboptimal concentration of thiamine for the growth. Iguchi et al.<sup>15)</sup> studying on the effect of antibiotics on the production using Corynebacterium petrophilum isolated by them, have also shown the accumulation of 13 g/l of glutamic acid from 3% g/l of n-hexadecane by addition of penicillin to the growing culture. Both the techniques, by limitation of the concentration of growth factor<sup>17)</sup> and by addition of penicillin<sup>18)</sup>, have been developed as excellent means for the production of this amino acid by the conventional fermentation process in which carbohydrates were used as the raw material.

During the course of the study on the optimal condition for glutamic acid production, Imada et al.<sup>19)</sup> have indicated that the yield of 11 g/l was obtained from 70 g/l of n-pentadecane without penicillin under the concentration of 0.025% of corn steep liquor. In this work they emphasized the effectiveness of iron ion in the culture medium for the basic that the production of glutamic acid was absolutely dependent upon the concentration of iron higher than 0.2 mg/ml, while in the lower concentration, the alcohol and fatty acid corresponding to the n-paraffin used above was easily utilized, but the oxidation of n-pentadecane was significantly depressed. They speculated from the results that iron may participate in oxygenase reaction in the earlier step of n-paraffin metabolism. In addition, it has been reported that the effectiveness of thiamine on this process was enhanced by the addition of small amounts of cysteine. Ueda et al.<sup>20)</sup> have found the effectiveness of cysteine as well as iron ion, and also indicated that the optimal concentration of phosphate was at the range of 0.5 to 1.0% in this process.

In connection with our recent work<sup>21)</sup> concerning luteinic acid production from n-paraffin using Aerobacter parvilineum and Corynebacterium sp. which also required thiamine for growth, it has been reported that under the sub-optimal condition of thiamine concentration, the yield of luteinic acid was about 30 g/l and that of N-aceto-butyric acid was approximately 50 g/l, from 100 g/l of n-paraffin mixture mainly containing  $\beta_{12}$  to  $\beta_1$  fractions. On the other hand, the addition of penicillin to the exponentially growing culture of the above strains under the optimal condition of thiamine concentration for their growth stimulated preferably the yield of luteinic acid and resulted in the production of more than 70 g/l, whereas the production of  $\alpha$ -ketobutyrate was depressed below 5 g/l. It has been obtained from the experiments of semi-practical scale using the jar-fermenter equipped with non-magnetic capable of controlling pH of the medium automatically by ammonia solution.

In view of these investigations, it might be possible that the process of glutamic acid production will make an industrial scale in the near future. Nevertheless, for the further progress of this process, it is necessary to combine systematically the studies from microbiological, biochemical and bioengineering view point.

### Lysine and other amino acids

Following the procedure of Saito<sup>14</sup> for the production of  $\alpha$ -paraffin as shown above, and after appropriate modification to find the microbe capable of producing citric acid, and the amount was adjusted, the quantities of amino acids produced were those shown in Table I. It has been reported that *Candida cylindracea* Difesa, *Candida cylindracea* produces methionine and lysine in the culture media.<sup>15</sup> In 1951, T. Kuroda et al.<sup>16</sup> in attempting to obtain auxotrophic mutants of *Candida cylindracea* for methionine, have isolated arginine-requiring mutants. *Candida cylindracea* H-302, he reported the production of methionine from paraffins and the yield was 1.4 or 6.2 g/l from 10% paraffin at 30°C. and off the substrate, 10 or 5% v/v. Oshima and Horio<sup>17</sup> in 1951, also reported the same auxotrophic auxotrophs which required the addition of 10% yeast extract or 1% fish meal. Arginine-requiring mutants of *Candida cylindracea* H-302 were obtained, they have isolated another auxotroph, *Candida cylindracea* H-302 which requires arginine as well and found that it can grow on  $\alpha$ -paraffin. From  $\alpha$ -paraffin mixture containing the fractions of 10% each of  $\alpha$ -paraffin, 10% yeast extract, 10% fish meal, 10% v/v from 10% v/v of yeast extract, 10% fish meal, 10% v/v yeast extract which requires methionine, the yield was 1.4 g/l reported by Yamagami et al.<sup>18</sup> in 1953. In addition, the same auxotrophs as isolated for *Candida cylindracea*, (10 to 50% v/v), 10% yeast extract, 10% fish meal, the yield was 1.4 g/l from 10% v/v yeast extract. On the other hand, *Candida*,<sup>19</sup> *Pseudomonas*,<sup>20</sup> *Paracoccus*,<sup>21</sup> *Acinetobacter*,<sup>22</sup> *Leptothrix*,<sup>23</sup> *Leptospirillum*,<sup>24</sup> *Leptothrix* and *Leptospirillum* are auxotrophic mutants. In *Candida*, it is difficult to find the auxotrophs for the utilization of hydrocarbon because of the considerable difficulty in finding the auxotrophic strain. Authors<sup>25</sup> in 1953, reported the close identity between propionity of amide nitrogen requiring auxotroph and the auxotrophs comparable to them in carbo-cyanoamino acids. The result is:

### 2. Application of $\alpha$ -paraffin and derivatives

Isocyanides and isocyanates are one of the structural structural materials in living organisms. Therefore, these microbes are capable of growing on hydrocarbons and they may have the energy metabolic system by which hydrocar-

bons are converted into sugars. Actually, it has been reported that some microorganisms grown with hydrocarbon reduced significant amounts of polysaccharide which presumably has the function of capsular material. Raymond et al.<sup>25)</sup> have shown that a polysaccharide was accumulated in the cultures of *Neocarina* grown with *n*-octadecane.

Recently, we reported the extracellular accumulation of glucose and trehalose by bacteria grown on *n*-paraffin as the sole source of carbon.<sup>26)</sup> Further, the addition of penicillin to the growing culture led to a remarkable increase in the accumulation of trehalose. The yield attained to approximately 3 g/l, corresponding to 60 to 70 % of total amount of sugar accumulated in the culture medium.

Of interest is that these sugars were also produced by other bacteria which can utilize *n*-paraffin as the sole source of carbon. However, it was explained that trehalose was derived from trehalose lipid which was present on bacterial cells and played a possible role in the utilization of *n*-paraffin by bacteria. Penicillin significantly suppressed the biosynthesis of this lipid and consequently led the extracellular accumulation of one of the precursors, trehalose and fatty acid.

We have also found that various polysaccharides were produced from *n*-paraffin by bacteria.<sup>30)</sup> For instance, arabomannan was isolated from the culture medium. Arabanose, mannose, galactose, glucose, rhamnose and unknown pentose were identified as constituents of the polysaccharide. Recently, it was found that yeast strain of Candida and Lachnomyces produced considerable amounts of mannan and unknown sugar from *n*-heptadecane culture<sup>31)</sup> that nonionic surfactant was supplied to the culture medium.

The study of sugar production from hydrocarbons is still in the early stage and it can be expected to develop further.

#### 4. Production of organic acid

According to the reported route of microbial oxidation of hydrocarbons<sup>32-34)</sup>, *n*-paraffin is firstly oxidized to fatty acids and is further converted to lower acids by the method either of  $\alpha$ - or  $\omega$ -oxidation. From this viewpoint, it appears to be possible to get a variety of intermediates by

this route. Actually, the production of dicarboxylic acid such as pimelic acid and adipic acid have been reported.

On the other hand, it has been published that a considerable amount of citric acid was produced by yeast strains on n-alkanes. Abe and his colleagues<sup>(35,36)</sup> who had isolated the yeast strain incapable of converting efficiently n-paraffin to citric acid, have shown that 30 g/l of citric acid was produced from 60 g/l of n-hexadecane through the fermentation process using Candida lipolytica, in which the elimination of iron ions out of the medium was essential for the increase in the rate of citric acid to isocitric acid produced.

We have also performed a study of citric acid production on a semi-industrial scale using mold and yeast strains, Penicillium panthinellum KY 1141, Candida cylindroides nov. sp KY 5002<sup>(37)</sup>. When the latter was grown aerobically on n-paraffin mixture mainly containing C<sub>12</sub> to C<sub>15</sub> fractions the yield of citric acid was more than 65 g/l, corresponding to 1.5% of n-paraffin used.

The accumulation of  $\alpha$ -keto-glutaric acid and pyruvic acid have been also reported using other yeast strain, Candida lipolytica AJ 5004, isolated by the research group of Ajinomoto Co. Ltd.<sup>(38,39)</sup>. The yields were respectively 60 g/l and 4 g/l from 10 g/l of n-paraffin as the carbon source. In this process, the effectiveness of calcium ion on the production of organic acids was emphasized.

We may also report the production of  $\alpha$ -keto glutaric acid by Arthrobacter paraffineus which was isolated as glutamic acid-producing strain as described above<sup>(5,21)</sup>. By adjusting pH automatically to 6.5 - 7.0 during the fermentation, more than 70 g/l of K<sub>2</sub>CO<sub>3</sub> to acid and 30 g/l of glutamic acid have been produced from 100 g/l of n-paraffin mixture under the suboptimal concentration of thiamine in the culture medium.

## 5. Production of vitamins and co-enzymes

Advantages claimed generally for fermentation processes are that many substances having chemically complex structures are obtainable more easily than by synthetic methods. The fermentation production of vitamins and co-

enzymes may be placed under this category.

Attempts to produce thiamine chalcocins by microbial methods from hydrocarbon has also been actively carried out in Japan.<sup>40)</sup> Suruki et al.<sup>41)</sup> have reported the accumulation of  $\text{B}_2$  in the culture medium of yeast strains belonging to Lichia. According to the cell growth, over the concentration of n-paraffin of 0.5 - 2.0 %, accumulating amount increased and the maximum yield reached approximately 51 mg/l. Sato et al.<sup>42)</sup> have also shown the production of riboflavin from n-hexadecane by Eremothecium ashbyii grown on the medium containing ammonium biphosphate, sodium glutamate, liquid paraffin and Tween 80. The yields approximately 9 mg/l, of which major part extracellularly accumulated.

Vitamin  $\text{B}_6$  which is indispensable for the oxiditive reaction of amino acids or nucleosides is known to be contained widely in microbial cells. When hydrocarbon was applied for the substrate, too, it has been reported<sup>40)</sup> that yeast such as Candida albicans and some of Lycobacterium produced this vitamin in their culture media. The yield was not higher than 400  $\mu\text{g}/\text{l}$ , but it is planned to be developed further.

Increasing attention has been paid towards deoxyadenosyl- $\text{B}_{12}$  since its biological function was recognized as the factor of antiperiodic chemi. Fukui et al.<sup>40)</sup> have attempted to produce it by Corynebacterium simplex and rocardia lutetiae reported the yield of 166  $\mu\text{g}/\text{l}$  from n-hexadecane by Corynebacterium simplex.

The quantity of these vitamins produced up to now is not enough to develop on a larger scale.

On the other hand, Tsukui et al.<sup>43)</sup> studying the production of biotin, has reported that a strain of Pseudomonas isolated from the soil produced about 30 mg/l of biotin- $\alpha$ -D-tet compound, characterized mostly as lesthisticin.

The attempts to produce carotenoids and cytochrome b' have been carried out as well. Nishimura et al.<sup>44)</sup> have shown that Lycobacterium sanguinis grown with kerosene as carbon source accumulated xanthophyll, of which the main components were 4-keto- $\gamma$ -carotene and its derivatives. Subsequently,

Fukui et al.<sup>40)</sup> have reported that the yield of xanthophyll was approximately 3 mg/l in a laboratory scale experiment. Besides, cytochrome 'c' has been produced by several yeast strains such as Candida glycinina and Torulopsis utilis. Under the condition, anionic surfactant was added, about 10 mg/l of cytochrome 'c' has been accumulated.

In addition, the bacterial production of coenzyme Q<sup>45)</sup> and coenzyme A<sup>46)</sup> have been reported by Fukui et al. in the research group of Takeda Pharmaceutical Company. The production of these compounds probably makes a further improvement, because these coenzymes participate in the sequence of oxidative reaction of hydrocarbon, whereby it is presumed to be contained in hydrocarbon-utilizing microorganisms much more than others.

#### 6. Production of other biochemical metabolites

Of the special materials derived from n-alkane through the microbial action, the production of phenazine and its derivatives have been shown. It has been reported that Phenazin-5-carboxylic acid was accumulated in the hydrocarbon layer of the culture medium<sup>47)</sup>. We have also observed that orange and purple-colored pigments were accumulated in a crystalline form in a n-paraffin layer of the culture medium of Arthrobacter perfringens KY 7134 and identified respectively as phenazine-1, (-hydroxide and phenazinediol-5, 10-N-oxides<sup>48)</sup>. There may be no reason for the reason why the rich compounds are producible from straight chain hydrocarbon.

Generally, during the growth of bacteria with hydrocarbon, it is often observed that the lysis of bacterial cells occurs with its growth. It is particularly noted when penicillin, cephalosporin C and its derivatives were added to the growing culture<sup>21)</sup>. Applying this technique, various cell components have been reported to be excreted to the culture medium. For example, protein<sup>49)</sup>, nucleic acids<sup>49)</sup>, UEPG peptide<sup>50)</sup>, phospholipid<sup>51)</sup>, and polysaccharide<sup>29)</sup> have been already shown.

#### 7. Production of chemicals from aromatic hydrocarbon

Comparatively, little is known about microorganisms which can utilize aromatic hydrocarbons as compared with aliphatic compounds. By the appli-

cation of co-oxidation phenomena, Raymond et al.<sup>52)</sup> have reported the microbial oxidation of aromatic compounds.

Follmann et al.<sup>53)</sup>, Datta and his coworkers<sup>54)</sup> have isolated the bacterium *Bacillus leonis*, *Bacillus licheniformis*, *Leuconostoc* sp., new aromatic hydrocarbon as the sole source of energy, and reported the modification of p-xylene or cumene (tri-oxo-cyclohexane, 75% min. yield) to cyclohexanone by *psuedomonas* sp. culture, while, Matsuura et al.<sup>55)</sup> found the reduction of 50 g/l of n-hololeic acid (max 20%), of 17% in the control group, by *Microbacter*.

The biological conversion of n-hololeic acid to behenic acid has already been known. Recently, Follmann et al.<sup>56)</sup> have succeeded in obtaining a yield of 12 g/l of behenic acid which is higher than the overall percent of 10% of *Microbacter* group, until now. In this process, acetyl acetone was used as a preferable co-substrate due to its strong uncoupling activity. Additionally, Follmann et al.<sup>57)</sup> have shown the possibility to improve the production yield through the use of yeast protein as a nucleophilic exchange resin for the esterification of behenic acid to behenic acid produced out of the fermentation system.

The conversion of behenic acid to behenic ester has been demonstrated by Follmann et al.<sup>58)</sup>. The esterification initiated by their were able to convert 10% behenic acid (10 g/l) to behenic acid (1.12 g/l).

Although the authors have not mentioned the utilization of aromatic compounds as sole carbon source, it is difficult to apply them directly to the synthesis of behenic acid. It is due to the difficulty in microbial oxidation of aromatic compounds, especially in synthetic activity, in comparison to the oil oxidation. Most the compounds in the microbial systems.

As mentioned, we have done some simple studies on the microbial production of various useful chemicals from cellulose and aromatic hydrocarbons. Besides, methanol has been proven to be working with the utilization of the same hydrocarbons derived from petroleum. Actually, ethanol<sup>59)</sup>, acetic acid<sup>60)</sup>, cellulose<sup>61)</sup> and so on, have been applied as the substrate for glutamic acid production.

As it has been described above, it is confident that the techniques of hydrogenation will make much progress in Japan. When full industrial operation is accomplished in the near future, we will be able to overcome the difficulties in all conventional processes where the supply of the raw materials is obliged to depend on agricultural products, and it will render great contribution to the prosperity of human beings.

We regret that some of the pages in the microfiche copy of this report may not be up to the proper legibility standards, even though the best possible copy was used for preparing the master fiche.

References

- 1/ "Elementary petroleum microbiology" ed. by J.M. Sharpley, Gulf publishing Co., Houston, Texas, 1975.
- 2/ "Petroleum microbiology" ed. by J.M. Davis, Elsevier Publishing Co., New York, 1967.
- 3/ T. Iwasa, S. Nakahashi, K. Yamada, K. Uchida and H. Kida, Amino acid and nucleic acid, ibid, 15 (1966)
- 4/ T. Iwasa, S. Nakayama and T. Takeda, ibid, 17, 136 (1968)
- 5/ K. Tanaka, K. Yamada, T. Suzuki, T. Yamaguchi and S. Kinoshita, J. Ferment. Techn., 47, 291 (1969)
- 6/ T. Takeda et al. S. Moriguchi, J. Ferment. Assoc. Japan, 26, 210 (1968)
- 7/ T. Furuta, ibid, 26, 1 (1968)
- 8/ S. Kuroda, Petroleum and Petrochemistry, Japan, 12, 64 (1968)
- 9/ N.A. Kirilova, Biotekhnika, Biophys., 1, 549, 567 (1956)
- 10/ T. Suzuki, K. Yamada, T. Matsubara and S. Kinoshita, Abstract of Paper, Annual meeting of the Jap. Chem. Soc. Japan, 1969, p 253.
- 11/ K. Yamada, J. Ferment. Assoc. Japan, 16, 37 (1968)
- 12/ T. Shioya, O. Otsuka, S. Hashimoto and H. Ueda, J. Gen. Appl. Microbiol., 19, 23 (1963)
- 13/ T. Iwasa and T. Nakaya, ibid, 1, 267 (1964)
- 14/ T. Iwasa, S. Nakayama, T. Yamaguchi and T. Takeda, Amino acid and nucleic acid, 7, 19 (1963)
- 15/ T. Iwasa, S. Nakayama and T. Takeda, ibid, 10, 86 (1966)
- 16/ T. Iwasa, K. Yamada, T. Nakai and T. Yamada, ibid, 10, 1 (1964)
- 17/ T. Iwasa, T. Yamada and S. Kinoshita, J. Agr. Chem. Soc. Japan, 34, 559 (1960)
- 18/ T. Iwasa and T. Nakaya, ibid, Patent, 593, 497 (1961)
- 19/ T. Iwasa, R.C. Pratkanew and T. Yamada, Amino acid and nucleic acid 16, 11 (1967)
- 20/ T. Yamada, S. Itoh and T. Shioya, Abstract of Paper, Annual meeting of the Jap. Chem. Soc. Japan, 1968, p 370.
- 21/ K. Yamada, T. Yamada, K. Timura, K. Tomita and S. Kinoshita, ibid, 12, 1, p 234.
- 22/ J.L. Dornan, Jr., J. Pankosa and R.L. Raymond, USP 3, 219, 543
- 23/ R. Imai, O. Otsuka and T. Shioya, J. Gen. Appl. Microbiol., 13, 217 (1967), ibid, 1, 105 (1967)

- 24/ K. Tanaka, K. Ochiai, T. Sekine and S. Kinoshita, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1968, p 160.
- 25/ Y. Suzuki, K. Ochiai, K. Tanaka and S. Kinoshita, Amino acid and Nucleic acid, 12, 165 (1969)
- 26/ Y. Suzuki, K. Ochiai and S. Kinoshita, Agr. Biol. Chem., 31, 121 (1967)
- 27/ K. Tanaka, Y. Suzuki, H. Imai, K. Ochiai and S. Kinoshita, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1969, p 185.
- 28/ K. Tanaka, T. I. Derry, J. Bacteriol., 129, 735 (1960)
- 29/ T. Suzuki, K. Tanaka and S. Kinoshita, Anal. Chem., 31, 190 (1969)
- 30/ K. Tanaka, Abstract of Paper, unpublished.
- 31/ K. Tanaka, Abstract of Paper, unpublished.
- 32/ K. Tanaka and T. I. Derry, J. Bacteriol., 129, 183 (1965)
- 33/ T. Suzuki, Abstract of Paper, Biochimica et Biophysica Acta, 24, 231 (1952)
- 34/ K. Tanaka, Abstract of Paper, Chem. Soc. Japan, 31, 727 (1967)
- 35/ K. Tanaka, T. I. Derry, Anal. Chem. Soc. Japan, 31, 727 (1967)
- 36/ K. Tanaka, T. I. Derry and S. Kinoshita, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1969, p 163.
- 37/ K. Tanaka, T. I. Derry, K. Ochiai and S. Kinoshita, unpublished
- 38/ K. Tanaka, T. I. Derry, K. Ochiai and O. Okamura, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1968, p 360.
- 39/ K. Tanaka, T. I. Derry, Anal. Chem. Soc. Japan, 31, 183.
- 40/ K. Tanaka, T. I. Derry, Anal. Chem. Soc. Japan, 26, 19 (1968)
- 41/ K. Tanaka, T. I. Derry and S. Kinoshita, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1968, p 163.
- 42/ K. Tanaka, T. I. Derry and S. Kinoshita, Vitamine, Japan, 34, 542 (1969)
- 43/ K. Tanaka, T. I. Derry and S. Kinoshita, Anal. Chem., 30, 1238, 1968 (1969)
- 44/ Y. Nishizawa and H. Yamada, Annual meeting of the Agr. Chem. Soc. Japan, 1969.
- 45/ K. Tanaka, T. I. Derry and S. Kinoshita, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1969, p 164.
- 46/ K. Tanaka, T. I. Derry, M. Nakano and S. Kinoshita, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1969.
- 47/ K. Tanaka, T. I. Derry, M. Nakano and A. Sato, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1969.

- 48/ T. Suzuki, T. Deguchi, K. Tanaka and S. Kinoshita, unpublished.
- 49/ T. Suzuki, K. Yamaguchi, K. Tanaka and S. Kinoshita, unpublished.
- 50/ S. Yamatoya, Y. Nakao, T. Kanazawa and A. Matsui, Symposium on Amino acid and Nucleic acid, Japan, 1963.
- 51/ M. Kikuchi, K. Sugiyama, A. Oki, H. Suzuki and Y. Nakao, Abstract of Paper, Annual meeting of the Agr. Chem. Soc. Japan, 1969, p. 185.
- 52/ J.B. Davis and R.L. Raynor, J. Appl. Microbiol., 2, 373 (1961)
- 53/ K. Yamada, S. Horiguchi and J. Takemoto, Agr. & Biol. Chem., 29, 943 (1965)
- 54/ T. Onori, S. Horiguchi and K. Yamada, ibid, 31, 1337 (1967)
- 55/ T. Ichikawa, H. Nishida, K. Fanno, H. Miyachi and A. Ozeki, Agr. Biol. Chem., 32, 12 (1968)
- 56/ H. Tone, S. Kitai, J. Ichizuka and I. Okaki, Annual meeting of the Ferment. Tech. Japan, 1967.
- 57/ K. Ogata and H. Asagi, Agr. Biol. Chem., 30, 116 (1966)
- 58/ T. Oki, Y. Suyama, Y. Nishimura and I. Onuki, Agr. Biol. Chem., 32, 119 (1968)
- 59/ T. Tsunoda, I. Shioi and K. Matsushige, J. Gen. Appl. Microbiol., 7, 30 (1961) K. Miyama, ibid, 10, 23 (1964)
- 60/ Miyagi, Matsui, Kitai, Tone and Tsunoda, Patent Journal, Japan, 37-9298



**26. 5. 72**